



Review Cancer neuroscience: State of the field, emerging directions

Frank Winkler,^{1,16,*} Humsa S. Venkatesh,^{2,16} Moran Amit,³ Tracy Batchelor,² Ihsan Ekin Demir,⁴ Benjamin Deneen,⁵ David H. Gutmann,⁶ Shawn Hervey-Jumper,⁷ Thomas Kuner,⁸ Donald Mabbott,⁹ Michael Platten,¹⁰ Asya Rolls,¹¹ Erica K. Sloan,¹² Timothy C. Wang,¹³ Wolfgang Wick,¹ Varun Venkataramani,^{1,8,16,*} and Michelle Monje^{14,15,16,*} ¹Neurology Clinic and National Center for Tumor Diseases, University Hospital Heidelberg and Clinical Cooperation Unit Neurooncology, German Cancer Consortium (DKTK), German Cancer Research Center (DKFZ), Heidelberg, Germany

²Department of Neurology, Brigham and Women's Hospital, Boston, MA, USA

³Department of Head and Neck Surgery, MD Anderson Cancer Center and The University of Texas Graduate School of Biomedical Sciences, Houston, TX, USA

⁴Department of Surgery, Technical University of Munich, Munich, Germany

⁵Center for Stem Cells and Regenerative Medicine, Baylor College of Medicine, Houston, TX, USA

⁶Department of Neurology, Washington University, St Louis, MO, USA

⁷Department of Neurosurgery, University of California, San Francisco, San Francisco, CA, USA

⁸Department of Functional Neuroanatomy, University of Heidelberg, Heidelberg, Germany

⁹Department of Psychology, University of Toronto and Neuroscience & Mental Health Program, Research Institute, The Hospital for Sick Children, Toronto, Canada

¹⁰Department of Neurology, Medical Faculty Mannheim, University of Heidelberg, Heidelberg, Germany

¹¹Department of Immunology, Rappaport Faculty of Medicine, Technion – Israel Institute of Technology, Haifa, Israel

¹²Monash Institute of Pharmaceutical Sciences, Drug Discovery Biology Theme, Monash University, Parkville, VIC, Australia

¹³Department of Medicine, Division of Digestive and Gastrointestinal Diseases, Columbia University, New York, NY, USA

¹⁴Department of Neurology and Neurological Sciences, Stanford University, Stanford, CA, USA

¹⁵Howard Hughes Medical Institute, Stanford University, Stanford, CA, USA

¹⁶These authors contributed equally

*Correspondence: frank.winkler@med.uni-heidelberg.de (F.W.), varun.venkataramani@med.uni-heidelberg.de (V.V.), mmonje@stanford.edu (M.M.)

https://doi.org/10.1016/j.cell.2023.02.002

SUMMARY

The nervous system governs both ontogeny and oncology. Regulating organogenesis during development, maintaining homeostasis, and promoting plasticity throughout life, the nervous system plays parallel roles in the regulation of cancers. Foundational discoveries have elucidated direct paracrine and electrochemical communication between neurons and cancer cells, as well as indirect interactions through neural effects on the immune system and stromal cells in the tumor microenvironment in a wide range of malignancies. Nervous system-cancer interactions can regulate oncogenesis, growth, invasion and metastatic spread, treatment resistance, stimulation of tumor-promoting inflammation, and impairment of anti-cancer immunity. Progress in cancer neuroscience may create an important new pillar of cancer therapy.

INTRODUCTION

As the nervous system governs such wide-ranging functions of the human body in health and disease, it is somewhat surprising that it took so long to fully appreciate its central involvement in cancer. Both the central nervous system (CNS) and the peripheral nervous system (PNS) regulate physiological functions and pathophysiological processes. Based on converging evidence, it is increasingly understood today that CNS activity and PNS activity regulate development, organogenesis, homeostasis, plasticity, regeneration, and immune function in diverse tissues (for review, see Boilly et al.¹ and Kumar and Brockes²). As cancer formation, growth, and progression subvert and repurpose mechanisms of development and regeneration, the nervous system may be implicated in all aspects of cancer pathophysiology. Reciprocally, cancer and cancer therapies can influence and remodel the nervous system, contributing to pathological feedback loops that not only yield neurological dysfunction but can also drive malignancy. These new insights have culminated in the emergence of cancer neuroscience as a new discipline³ that focuses on defining and therapeutically targeting nervous system-cancer interactions, both in the local tumor microenvironment and systemically.

In this review, we will provide an update on the current state and future directions of cancer neuroscience. We identify important unanswered questions and current roadblocks, specifying ways to overcome these obstacles through the implementation of cross-disciplinary development of technologies, knowledge, and scholarly infrastructure. Reciprocal interactions of cancers







Figure 1. Mechanisms of nervous system-cancer interactions

The nervous system (gray) and cancer (red) interact in at least six ways.

(A) Electrochemical interactions, including bona fide neuron-to-cancer synapses.

(B) Paracrine interactions from neurons/nerves to cancer cells, directly or through signaling with cells in the tumor microenvironment (green stromal cell and red blood vessel shown). In turn, cancer cells often secrete signaling molecules such as synaptogenic factors or axonogenic factors that locally remodel the nervous system to augment nervous system-cancer interactions.

(C) Systemic nervous system-cancer interactions, e.g., circulating neurotransmitters or neuropeptides that can influence cancer pathogenesis directly or indirectly such as through altered immune system (blue) function. Reciprocally, cancers can influence the nervous system at a distance through circulating factors or altered afferent neural signals.

(D) Three-way interactions between neurons or nerves, cancer cells, and immune cells can modulate anti-cancer immunity and pro-cancer inflammation. (E) Cancer cells may leverage cell-intrinsic signaling and other processes classically associated with neural cells. For example, autocrine neurotrophin signaling is

illustrated.

(F) Cancer therapies (chemotherapy, green) can profoundly alter nervous system function, including impaired function of various types of peripheral nerves and impaired cognitive function.

with the nervous system are discussed, with new multidisciplinary research subfields like "neuro-immuno-oncology" outlined. Importantly, a roadmap for clinical translation is laid out for the implementation of neuroscience-instructed cancer therapies. We make the case that cancer neuroscience (Figure 1) can stimulate both fields: cancer research and clinical oncology, as well as neuroscience and neuro-medicine, with synergy at the intersection of these disciplines.

Impact of the nervous system on tissue development, homeostasis, and plasticity

Neuronal activity influences organ development, homeostasis, plasticity, and regeneration—both in the CNS and throughout the entire body. The cellular and molecular basis for neuronal activity-dependent regulation of physiology in health has the po-

tential to provide insights into how the nervous system might similarly influence tumor biology. Given how instructive understanding development of the brain itself has been for the study of cancer neuroscience, we begin with an in-depth discussion of nervous system development to explore foundational concepts mirrored in cancer pathogenesis discussed later.

Central nervous system

Development of the CNS involves coordinated neuronogenesis and gliogenesis from neural stem and precursor cells; diversification of these neurons, astrocytes, and oligodendrocytes; migration of new cells to the appropriate location; and neural circuit assembly (for review, see Silbereis et al.⁴). Functional neural circuit development requires axonal outgrowth and pathfinding, establishment of synapses, and refinement of these connections between neurons. Astrocytes promote synaptogenesis, develop







a gap junction-coupled network throughout the brain, and engage with synapses to support synaptic function, while oligodendrocytes myelinate axons to provide metabolic support⁵ and enable fast saltatory conduction of action potentials.⁶

Electrical activity influences all aspects of nervous system development (for review, see Spitzer⁷). During early stages of neurodevelopment, synchronous waves of electrical activity and consequent voltage-dependent calcium transients occur in developing neural tissues and regulate both cellular and synaptic patterning. In the nascent brain, gap junctions couple neural stem cells in the germinal zone, allowing membrane depolarization-induced calcium transients to propagate synchronously through the germinal zone, regulating stem cell proliferation.⁸ Early in neurodevelopment, neurotransmitters are secreted from a variety of cell types in a non-synaptic manner to promote the generation of neurons.⁹ Electrical activity also regulates the migration of these newly generated neurons¹⁰ and influences axonal pathfinding and axonal targeting.^{11–13}

In the developing nervous system, gap-junctional coupling occurs between migrating neuroblasts,¹⁴ neurons in the prenatal and early postnatal neocortex,^{15,16} and between neurons in numerous additional neuroanatomical locations. Such coupling, together with mechanisms of cell depolarization such as nonsynaptic glutamate secretion and "pacemaker" neurons,¹⁷ enables synchronized calcium transients to spread through developing CNS structures such as the nascent neocortex.¹⁸ Recent work has suggested that a small, distinct subpopulation of single neurons arborizes throughout the entire brain to provide a specific periodic signal coordinating brain development.¹⁹ Such experience-independent, coordinated waves of activity promote the assembly of functional neural circuits that are later refined in an experience-dependent manner.^{20,21}

Neurotransmitter signaling regulates brain organogenesis and later serves as the backbone of synaptic communication between neurons. The formation of new neurons from stem and progenitor cells, as well as their integration into neuronal circuits, is driven by neurotransmitters during development (for review, see Ojeda and Ávila²²) and in neurogenic regions of the adult brain.⁹ This signaling is fine-tuned and can be spatially and temporally hetero-

Figure 2. Parallel mechanisms of glial plasticity and glial malignancy

Left: neuron (gray) to oligodendroglial (blue) interactions involve neuron-to-oligodendrocyte precursor cell synapses and paracrine (red circles) signaling, e.g., BDNF-TrkB signaling, during development and throughout life. Neuronal activity can promote the proliferation of oligodendrocyte precursor cells, generation of new oligodendrocytes, and adaptive changes to myelination that tune neural circuit function. Such plasticity of myelin contributes to healthy cognitive function throughout life. Right: neuron to glioma (green) interactions involve neuron-to-glioma synapses and paracrine signaling, e.g., BDNF-TrkB signaling. Glioma hijacking of mechanisms that normally support myelin development, homeostasis, and plasticity instead contribute to glial cancer initiation, growth, and invasion.

geneous: for example, during development, the neurotransmitter GABA (which is an inhibitory neurotransmitter in later life) is chiefly excitatory (depolarizing) due to developmental expression patterns of chloride transporters and is implicated in many processes of neural development, including neuronal proliferation, migration, differentiation, and preliminary circuit-building in the CNS (for review, see Ojeda and Ávila²²), while inhibiting the generation of neuronal progenies from embryonic stem cells and peripheral neural crest cells during early embryogenesis.²³

Cellular plasticity in the CNS does not end at the time of birth or during childhood. As is the case during development, neuronal activity also governs ongoing cellular plasticity throughout life. Neuronal activity and neurotransmitter signaling robustly regulate the proliferation of neural precursor cells, including oligodendrocyte precursor cells (Figure 2),²⁴ and neural stem cells in the subventricular zone^{25,26} and hippocampus.^{27,28} Neuronal activity drives one of the most important features of plasticity and adaptation in the adult brain: ongoing generation and remodeling of myelin^{24,29,30} (Figure 2), which contributes to motor function,²⁴ motor learning,³¹ attention and short-term memory,³² memory consolidation,^{33,34} and social function.^{35,36} In health, adaptive myelination appears to be highly and specifically regulated, with precise circuit-specific and neuron subtype-specific^{24,37} activity-regulated changes in myelin that tune circuit dynamics to promote coordinated circuit function.^{33,38,39}

Neurons communicate with neural stem cells and progenitor cells by activity-dependent paracrine factors such as brainderived neurotrophic factor (BDNF)³² and by synaptic communication (Figure 2). Synaptic signaling is well established for oligodendrocyte precursor cells (OPCs), which receive synaptic input via glutamatergic (calcium-permeable AMPA receptormediated) and GABAergic (GABA_A receptor-mediated) neuronto-glial synapses.^{40,41} Such neuron-to-OPC synapses are unidirectional, with the OPC always in the postsynaptic position, and can be of a transient nature,⁴² which is compatible with rapid migration of OPCs. Synaptic input to OPCs is extensive, involving both short-range and long-range inputs,⁴³ although the role that such neuron-to-OPC synapses may play in activity-regulated myelination remains incompletely understood.



Beyond development and plasticity, glutamatergic neuronal activity also promotes myelin regeneration after a demyelinating injury.^{44,45} GABAergic signaling to OPCs is involved in resistance to and adaptive repair of hypoxia-induced dysmyelination.⁴⁶ Remarkably, new evidence suggests that following injury, not only neurons from the PNS but also those from the CNS can revert to an embryonic-like growth state that allows axonal regeneration.⁴⁷ Together, this speaks for a remarkable ability of the CNS to self-repair damage, at least to a certain extent, by neuronal activity-regulated mechanisms.

Neural activity is also an important regulator for vascular homeostatic physiological processes in the CNS. One example is the neural auto-regulation of cerebral blood flow in the brain, called neurovascular coupling, which allows regional blood flow to increase to quickly supply oxygen and nutrients according to demand⁴⁸; this process involves neurons, astrocytes and vascular cells, and includes direct neurotransmitter signaling.⁴⁹ Neuronal activity also directly regulates the blood-brain barrier by modulating endothelial gene expression and the functions of efflux transporters.⁵⁰

Peripheral nervous system

Innervation similarly regulates tissue development, organogenesis, and regeneration outside the CNS, throughout the entire body (for reviews, see Boilly et al.¹ and Kumar and Brockes²). The CNS controls a myriad of non-neural cells and bodily functions, either by hormone secretion into the systemic circulation or in a more region-specific manner via the PNS, which connects the CNS to all organs via sympathethic (adrenergic), parasympathetic (cholinergic), motor, and/or sensory nerve fibers.

The role of nerves in development is increasingly appreciated, with organogenesis depending on proper innervation. A strong dependence on functional nerves and undisturbed nerve growth is long known to be indispensable for limb regeneration in amphibia and reptiles.⁵¹ In mammals, a similar dependency of organogenesis on innervation has been reported. For the example of the salivary gland, parasympathetic innervation is crucial for glandular organogenesis.⁵² Likewise, heart regeneration in neonatal mice is impaired by denervation,⁵³ and heart organogenesis depends on sympathetic nervous system signaling.⁵⁴ It is an exciting question to address whether synapses, or synapse-like structures, exist between neurons and certain non-neuronal cells throughout the body. The answer to this question also has great implications for cancer neuroscience and would help elucidate neuron-tumor interactions in the light of neurodevelopmental processes.

The innervation of tissue stem cell niches also regulates the functions of various cell types, both during development and in mature tissue, as demonstrated for the skin,^{55–57} gastrointestinal tract,⁵⁸ and bone marrow.⁵⁹ Additionally, Schwann cells, the chief glial cell type of the PNS, are involved in maintenance of hematopoietic stem cells in the bone marrow niche.⁶⁰

The nervous system contributes to tissue regeneration throughout the entire body. Injured adult organs do not regenerate after denervation, while restoring the function of cholinergic signaling in salivary gland tissue improves epithelial regeneration.⁶¹ Likewise, epidermis regeneration during wound healing depends on nerve-derived sonic hedgehog signaling, allowing hair follicle stem cells to become epidermal stem cells.⁵⁶

As discussed in detail below, the nervous system is also involved in the regulation of multiple functions of the immune system (for review, see Schiller et al.⁶²). Moreover, nerves control blood vessels in the periphery: during development and tissue repair, blood vessels and nerves use similar signals and principles to differentiate, grow, and navigate toward their target. The release of sympathetic neurotransmitters has been implicated in the formation of new blood vessels during these processes.⁶³ Furthermore, sympathetic innervations of the vessels can affect the extravasation of immune cells from the blood vessels to the local tissue by modulating their expression of adhesion molecules,⁶⁴ thereby affecting the local immune response.

In summary, the CNS and PNS are not only involved in cognitive functions, movement, and sensation, but they also govern the generation, adaptation, plasticity, and repair of tissues and organs. Local (paracrine) and systemic neural factors, classical synaptic contacts between neurons, as well as *bona fide* synaptic contacts to cells that are not mature neurons are involved in this complex, multilayered system of governance. This explains why neural-cancer interactions are so intriguing to study, since all of the "non-canonical" biological functions of neuronal activity described above are highly relevant for cancer as well: i.e., organo(/tumoro)genesis; growth by activation of developmental programs; invasion and colonization; control of a permissive microenvironment, including blood vessels and the immune system; and resilience and self-repair capabilities.

CNS cancer neuroscience

Paracrine signaling in brain tumor growth and initiation

As described above, neuronal activity controls vast and varied physiological functions. In parallel, nervous system activity and neural mechanisms can control brain tumor initiation, growth, invasion, and metastatic colonization of the brain. The idea that neurons may play a key role in brain tumor biology was first suggested by histological co-localization studies in 1938,⁶⁵ and application of the tools of modern neuroscience to study glioma biology has now demonstrated clearly that neuronal activity can drive brain cancer growth⁶⁶ (Figure 2). Mechanisms of activityregulated paracrine signaling were first appreciated with the discovery that neuronal activity-dependent paracrine signaling of neuroligin-3 (NLGN3), BDNF, and GRP7866-68 promote glioma proliferation and growth. Recent data show how even CNS tumor initiation can be driven by neuronal activity.⁶⁷ In addition to promoting glioma growth, NLGN3 regulates the initiation of optic gliomas in a cancer predisposition syndrome.⁶⁷ Activity-dependent shedding of NLGN3 is mediated by the metalloprotease ADAM10, and the growth of high-grade and low-grade gliomas were significantly decreased with ADAM 10 inhibitors in mouse models.^{67,69} Recently, IGF-1 was identified as another neuronal activity-regulated paracrine signaling molecule, which mediates olfactory sensory experience-dependent initiation of olfactory bulb high-grade glioma.⁷⁰ Together, these discoveries suggest that circuit-specific neuronal activity-dependent paracrine signaling differentially influences the neurobiology of distinct brain tumor types. Synaptic connections between neurons and brain

tumor cells

Brain tumor cells can structurally and electrically integrate into neural circuits. Accordingly, tumor cells from various adult and

pediatric glioma types form bona fide glutamatergic synapses with neurons (Figure 1A), driving tumor growth^{71,72} and brain invasion.⁷³ These synaptic connections consistently form unidirectionally from neurons on the presynaptic side to glioma cells on the postsynaptic side, inducing excitatory postsynaptic currents (EPSCs) predominately mediated by calcium-permeable AMPA receptors (AMPAR) in glioma cells.^{71,72} These EPSCs are depolarizing, and direct optogenetic depolarization of glioma cells increases glioma cell proliferation.⁷¹ Furthermore, inhibiting AMPAR function genetically or pharmacologically with perampanel, an FDA-approved anti-epileptic drug, reduces glioma cell proliferation and invasion.71-73 As discussed above, oligodendrocyte precursor cells (OPCs), a likely cell of origin for many types of glioma, and immature neurons also receive synaptic input,^{10,40} demonstrating that physiological correlates of malignant synaptic contacts exist.

Distinct from direct, *bona fide* synaptic interactions, indirect, perisynaptic contacts—reminiscent of the position an astrocyte normally assumes in a tripartite synapse—were found in breast cancer brain metastases⁷⁴ as well as adult glioblastoma.⁷² In breast cancer brain metastatic disease, glutamatergic signaling via these perisynaptic structures promotes tumor growth through NMDA receptors on the breast cancer cells.⁷⁴

Linking paracrine and synaptic mechanisms, NLGN3 induces a synaptogenic gene expression profile in glioma cells, which suggested it may act as an upstream regulator of malignant synaptogenesis.⁶⁹ Indeed, fewer neuron-to-glioma synapses form in the absence of NLGN3 in the tumor microenvironment.⁷¹ Paracrine BDNF signaling also promotes synaptic connectivity between neurons and glioma cells, as well as regulates the strength of malignant synapses.⁶⁸ Similar to the plasticity at physiological synapses that supports learning and memory in the healthy brain, glioma cell surface AMPA receptor trafficking is increased by BDNF, highlighting a postsynaptic mechanism of malignant synaptic plasticity.⁶⁸ In turn, this mechanism amplifies glutamate-induced inward currents in glioma cells and subsequently increases calcium transients. In patient-derived glioma cells, genetic or pharmacological inhibition of NTRK2 (BDNF receptor TrkB) consistently reduces glioma cell responsiveness to glutamate, decreases neuron-to-glioma synaptic connections, and reduces neuronal activity-induced glioma proliferation.68 Accordingly, pharmacological targeting of TrkB signaling in glioma inhibits tumor growth in mouse models without TrkB fusions,⁶⁸ highlighting a potentially broader indication for Trk inhibitors than only for gliomas expressing Trk fusions.

Brain tumor-induced modifications of the neuronal environment

Several mechanisms have been identified by which gliomas influence their neuronal microenvironment. Seizures caused by neuronal hyperexcitability are frequent in gliomas and brain metastases. Several paracrine factors and aberrantly increased neuronal synaptogenesis contribute to glioma-induced neuronal hyperexcitability. Paracrine glutamate secretion via the xc-cystine-glutamate transporter system increases neuronal hyperexcitability as well as glioma growth in models of adult glioblastoma.⁷⁵ In the tumor microenvironment of IDH-WT adult glioblastoma, loss of GABAergic interneurons also contributes to circuit hyperexcitability,⁷⁶ as does glioma-induced alterations

in neuronal chloride transporter expression, changing the effects of GABA from inhibitory to excitatory.⁷⁶ Another interesting mechanism promoting neuronal hyperexcitability is the ability of glioma cells to promote synaptogenesis, mirroring a physiological role of astrocytes.⁷⁷ In gliomas with specific point mutations of the enzyme PIK3CA, glioma cells secrete glypican-3 that drives aberrant synaptogenesis and associated neuronal hyperexcitability in mouse models,⁷⁸ indicating that distinct genomic characteristics of glioma can differentially affect the neuronal tumor microenvironment. Furthermore, gliomasecreted thrombospondin-1, another synaptogenic factor, promotes increased functional neuronal connectivity between the tumor and the brain; such functional connectivity of the tumor was strongly associated with decreased survival in humans with glioblastoma.⁷⁹

Taken together, these data highlight a positive feedback loop between neuronal hyperexcitability, neuron-glioma interactions, and brain tumor progression. This concept is strengthened by recent clinical data linking preferentially active brain regions to glioma occurrence.⁸⁰

Tumor-autonomous neurodevelopmental and neural mechanisms in brain cancer biology

In addition to neuron-tumor networks, brain tumor cells themselves show multiple neural and neurodevelopmental features, including network structures (Figure 1E). Ultra-long, neurite-like membrane protrusions called tumor microtubes (TMs) are used by glioma cells to scan the brain microenvironment,⁸¹ invade into the brain, 73,81-83 and colonize it by invasion and cell division.⁸¹ Over time, TMs interconnect single glioma cells to a functional, communicating multicellular network.73,81 TMs and the multicellular networks they generate are consistently found in human gliomas investigated so far, including astrocytomas grade 2-4 (which includes grade 4 glioblastomas), and K27Mmutated midline gliomas.71,72,81-85 As mentioned, many similarities exist between TMs and neural protrusions. A subpopulation of invasive TMs exhibits tips resembling the growth cones of neurites, neuronal processes during neurodevelopment that are essential for neuronal migratory pathfinding and network building.81,82 In addition, invasion-related features of TMs such as branching, protrusion, and retraction mimic mechanisms of neurite pathfinding.⁷³ Several molecular drivers of TM growth are also involved in neurite outgrowth and neurodevelopment, such as GAP-43 and TTYH1.81,83

Using gap junctions (mainly connexin 43) and adherens junctions between TMs, tumor cells interconnect with one another, building the anatomical basis of the tumor-tumor network. The network of tumor cells connected by gap junctions communicates via intercellular calcium waves and exchanges small molecules with one another, similar to physiological astrocyte networks in the brain.^{71,81,86} Importantly, this functional tumortumor network is a crucial factor for mediating therapeutic resistance. TM network-integrated, gap junction-coupled tumor cells were predominately resistant to radiotherapy and standard chemotherapy with temozolomide. By contrast, unconnected glioma cells were much more responsive to cytotoxic therapeutic treatment, which was associated with decreased tumor cellular homeostasis.^{81,85,87,88} This resembles mechanisms of normal brain astrocyte networks that can dilute toxic metabolites



throughout their gap junction-coupled network.⁸⁹ Furthermore, tumor cell coupling via gap junctions and TMs does not only occur with each other, but also with the astrocytic network of the brain, which has also been demonstrated for cancer cell survival in the brain during metastasis.^{73,90}

By contrast, glioblastoma cells not (yet) integrated into tumortumor or tumor-astrocyte networks are the drivers of glioblastoma invasion.⁷³ On a molecular level, this subpopulation was enriched for OPC-like, neural progenitor (NPC)-like and neuronal-like cell states. Interestingly, the invasive glioblastoma cell subpopulation showed migration patterns resembling immature neurons during neurodevelopment. Furthermore, analogous to immature neurons and OPCs receiving synaptic input, glioma cell invasion as well as TM dynamics and TM genesis were increased after neuronal stimulation.⁷³

In summary, while glioma cells that are connected with one another mediate therapeutic resistance, those that are not connected with one another or with astrocytes drive brain invasion. In other words, distinct neural features govern the various central traits of malignancy of aggressive brain tumors.

It has recently been discovered that TM-connected glioblastoma cell networks are characterized by autonomous rhythmic activity that is generated by pacemaker-like tumor cells. Residing in the hubs of the functional tumor networks, autonomously rhythmic tumor cells effectively influence the other network members via the generation of intercellular Ca²⁺ waves that travel throughout the network.⁹¹ In addition to neuron-to-glioma synaptic signaling that also generates Ca2+ activity, including Ca²⁺ waves in the glioma networks,^{71,72} this periodic activity is an alternative, tumor-autonomous mechanism of glioma network activation. Importantly, glioblastoma growth and cellular survival depended on this autonomous rhythmic activity, possibly via frequency-specific upregulation of distinct tumorpromoting intracellular pathways.⁹¹ Relevant for the field of cancer neuroscience, these findings show striking similarities to the spontaneous periodic network activity driven by pacemaker-like neuronal cells during neurodevelopment: regarding frequencies, molecular mechanisms for pacemaking (Ca2+-modulated potassium conductance), importance for network development, coordination of population activity, and plastic and even "self-repairing" features of pacemaker-like behavior.¹⁷ It will be interesting to learn whether other tumor types show a similar pathobiological mechanism by recapitulating this physiological neurodevelopmental principle.

The complexity of interactions between various components of tumors and the CNS (Figure 3) illustrates an important challenge for the future. In addition to the various neural mechanisms governing brain cancer biology, research in ion channels expressed in tumor cells, neural-tumor co-regulation of the blood-brain barrier, and tumor blood vessel biology, as well as other lines of research will certainly extend our knowledge in brain tumor cancer neuroscience.

PNS cancer neuroscience

Beyond the CNS, a wealth of studies across various cancer types has now demonstrated a fundamental role for the nervous system in driving tumor pathogenesis in cancers outside of the brain. As with gliomas, pathologists have appreciated the structural relationship between neurons and malignant cells in the periphery for more than a hundred years,⁹² largely due to the histopathological observances of perineural invasion (PNI)⁹³ that suggest that the perineural niche may be functionally beneficial to the tumor. PNI involves malignant cells surrounding or invading into nerve tracts, and it has been associated with aggressiveness and poor prognosis in a number of different cancers, including pancreatic, breast, and prostate cancers.^{94–96} As with gliomas, cancer cells of various non-CNS tumors have been found to display distinct neurodevelopmental features, at least on the gene expression level.⁹⁷

Preclinical studies have demonstrated an important role for the autonomic nervous system in the neural regulation of a wide range of cancers. For instance, in prostate cancer, *β*-adrenergic signaling (sympathetic) was found to be integral to tumor initiation, while cholinergic signaling (parasympathetic) contributed to invasiveness and dissemination.98 In breast and ovarian cancer, β-adrenergic signaling was found to accelerate cancer progression.99,100 Importantly, much like their differing roles in various tissues during normal development, different neuronal subpopulations may play distinct roles dependent on tissue type. As an example, cholinergic signaling has been shown to be either growth-promoting in gastric cancer, 101,102 or growth-inhibiting in pancreatic cancer.¹⁰³ Even within specific tissues, careful attention must be given to identifying the specific contributions of various neurotransmitters stemming from either parasympathetic or sympathetic nerve activity. For instance, in breast cancer, genetic manipulation of autonomic nerves revealed that sympathetic nerves accelerated tumor progression and growth, whereas parasympathetic nerves had the opposite effect.¹⁰⁴ Similarly, in pancreatic cancer, cholinergic signaling suppressed growth,¹⁰³ whereas adrenergic signaling promoted growth.¹⁰⁵

Sensory nerves have also been shown to play a role in cancer pathogenesis. Basal cell carcinomas require hedgehog signals from cutaneous mechanosensory sensory nerves for tumor formation,⁵⁷ and pancreatic cancers exhibit slowed growth with the ablation of sensory neurons.¹⁰⁶ In the context of metastasis, surgical denervation studies excluded a role for circulating catecholamines in stress-induced metastasis in a mouse model of breast cancer,¹⁰⁷ although sensory nerve innervation enhanced triple-negative breast cancer invasion and metastatic spread.¹⁰⁸ Thus, the specific impact of various neurotransmitters coming from the activity of different branches of the nervous systems on malignant tissues of all types must be carefully parsed (potentially even on a single-cell/cellular subpopulation-specific level) to better understand how manipulation of these neural circuits may be harnessed for treatment.

In the NF1 cancer predisposition syndrome, children and adults are prone to the development of benign peripheral nerve sheath tumors (neurofibromas) that derive from preneoplastic *NF1*-deficient Schwann cell precursors. These tumors are intimately associated with nerves, raising the intriguing possibility that neurons influence neurofibroma formation or growth. To this end, *Nf1*-mutant dorsal root ganglion neurons, which extend sensory axons to neurofibromas, exhibit greater action potential firing rates relative to wild-type controls. These *Nf1*-mutant sensory neurons also exhibit increased expression of collagen 1a2 that serves as a mitogen for *NF1*-deficient human and mouse







Figure 3. Therapeutic opportunities at the intersection of neuroscience and cancer biology

Increased understanding of nervous system-cancer crosstalk is beginning to elucidate therapeutic targets for a variety of cancers. While these targets vary in a tumor-specific manner, examples are shown here of the target structure or principle (green), a relevant molecular target (yellow), and of a drug or drug class that may prove useful for therapy (orange). Please note that only examples are shown, and each target is not necessarily relevant for every tumor type; for instance, targeting AMPAR-mediated synapses using the anti-seizure medication parampanel has to date only been demonstrated as a potential strategy for gliomas. Each therapeutic strategy requires testing in prospective clinical trials, which has been initiated for several of these examples (see text).

Schwann cells, such that inhibition of their excitability with the voltage-gated sodium channel blocker tetrodotoxin (TTX) or the anti-seizure drug lamotrigine reduces collagen 1a2 production as well as the growth of neurofibromas in *Nf1*-mutant mice *in vivo*.¹⁰⁹

Additional mechanisms promoting nerve-cancer interactions the tumor microenvironment include secreted neurotrophins that may be released in both activity- and non-activity-dependent manners from nerves or may be secreted from tumor cells. These neurotrophins, known to play a vital role in axonogenesis and nerve recruitment, have now been shown to critically modulate tumor growth outside of the brain (Figure 1). In pancreatic cancers, glial-cell-line-derived neurotrophic factor (GDNF)¹¹⁰ and artemin (ARTN)¹¹¹ secretion promotes PNI, while nerve growth factor (NGF) has been shown to recruit sensory nerves into the tumor microenvironment.^{112,113} Similar to gliomas, BDNF/NTRK signaling has been implicated in promoting tumor survival in multiple myeloma and ovarian cancers.^{114,115} Neuro-trophins are often upregulated by neural signaling through a

feedforward mechanism, with cholinergic signaling promoting NGF expression in gastric cancer¹⁰¹ and adrenergic signaling promoting NGF expression in pancreatic cancer¹⁰⁵; the NGF-induced increased nerve ingrowth into the tumor microenvironment further promotes tumor progression. Another avenue of neuronal contributions to the microenvironment of extracranial tumors includes metabolic support. Work by Zahalka and colleagues illustrated that β -adrenergic receptor signaling is critical for an angio-metabolic switch that fuels prostate cancer growth.¹¹⁶ In another example, pancreatic cancer cells increase NGF production to fuel metabolism.¹¹⁷

As illustrated in the above examples, in addition to benefiting from these secreted metabolites and neurotrophins, cancers reciprocally affect the nervous system (Figure 1). Just as brain tumors induce neuronal hyperexcitability, cancers outside of the CNS can increase innervation of the local tumor microenvironment by recruiting new nerve fibers via axonogenesis, ¹¹⁸ often driven by neurotrophin secretion. Another interesting example



of tumor-induced modulation of the nervous system that in turn fosters cancer progression has been suggested for prostate and other cancers where peripheral tumors attract doublecortin-expressing NPCs that leave the brain and home to the tumor via the blood stream, generating new neurons in the tumor which has growth-stimulatory effects.¹¹⁹ Remodeling of the neural microenvironment is further evidenced in a recent study demonstrating that tumor-associated neurons are reprogrammed toward an adrenergic phenotype that can stimulate tumor progression in oral cancer.⁹⁴ Together, these studies suggest that whether through activity-dependent mechanisms, paracrine signaling, or metabolic support, crosstalk between nerves and malignant cells (Figures 1B and 1C) in several tissues represents a novel angle to target malignant disease progression.

The ability of metastatic cells to leave the primary tumor and establish metastases is a major cause of death and a serious impediment to successful therapy. In brain metastases, these non-brain-cell-derived cancers hijack mechanisms of neurodevelopment (Figure 1E) for growth, as described above. Even outside the context of specific brain metastases, ion channels have been implicated in the overall metastatic process. Changes in potassium channel expression were found to alter metastatic breast cancer progression.¹²⁰ Recent studies have also more broadly suggested that a single ion channel, sodium leak channel, non-selective (NALCN), may regulate malignant cell dissemination and metastasis in a number of cancers.¹²¹ Investigating the broader role of neural activity in driving the metastatic cascade will also be critical as innervation of peripheral tumors has been linked to invasion and dissemination from primary tumors.⁹⁸ For example, sympathetic neural signaling through β-adrenergic receptors on breast cancer cells induced cvtoskeletal changes and protease production that increased breast cancer cell invasion.¹²² Sympathetic/β-adrenergic signaling to blood and lymphatic vessels in tumors contributes to metastatic dissemination.99,108 Together, these studies suggest that ion channel and neurotransmitter signaling in malignant cells and the tumor microenvironment may facilitate metastatic progression. Furthermore, a dietary-induced pro-regenerative state of peripheral glial cells (Schwann cells) was related to increased tumor innervation and metastatic potential.¹²³ In the future, studies elucidating the interactions between various types of neurons/ nerves and various types of metastatic cancers might lead to new strategies to prevent and treat metastatic spread. It will also be fascinating to learn whether metastatic cells become functionally integrated into neural networks, such as in glioma.

In summary, the distinct mechanisms of interactions between malignant cancer cells and neurons in their microenvironment are now being studied across different tissue types and organs, although much is yet to be understood about how peripheral cancers may integrate into neural networks and might respond to electrochemical neurotransmission. There is clear evidence that in oral squamous cell, head and neck, gastric, colon, rectal, prostate, breast, and pancreatic cancers, neurons of different types contribute to malignant tumor growth. Moving forward, evaluating the effects of direct activity-mediated neurotransmission to and membrane depolarization of these malignant cells will be an exciting area of study. New technologies that allow for interrogation, visualization, and quantification of neuronal activity within peripheral tumors will be needed (Figure 4) to unravel the neural inputs and signaling patterns that contribute to tumor pathogenesis. As this field evolves, it is imperative that all axes of neuronal communication with both neoplastic and nonneoplastic cells of the tumor microenvironment are thoroughly investigated.

Neuro-immuno-oncology

Neural cells respond to immune system signaling molecules, and immune cells respond to neurotransmitters and neuromodulators, so it is not surprising that neural-immune crosstalk can profoundly modulate both nervous system function and immune system function. In the context of cancer, a triangular relationship between neurons, immune cells, and cancer cells (Figure 1D) is emerging that is relevant to nervous system influences on the tumor immune microenvironment, pro- and anti-tumor immunity, and immunotherapy.

The autonomic nervous system plays key roles mediating communication between the brain and immune system. Afferent fibers of the vagus nerve convey information about peripheral immune challenges to the brain, and efferent vagus pathways modulate the immune response through cholinergic signaling, for example, powerfully mitigating pro-inflammatory cytokine release in the context of experimental lipopolysaccharideinduced sepsis.¹²⁴ Such an "inflammatory reflex¹²⁴" helps to exert precise control of powerful immune responses. This antiinflammatory influence of parasympathetic nerves and acetylcholine on peripheral immune responses is one such mechanism of control, while neural orchestration of immune cell trafficking and function by the sympathetic nervous system is another important mechanism of regulation. Adrenergic signaling via sympathetic innervation regulates physiological, diurnal trafficking of lymphocytes through lymph nodes¹²⁵ and egress of hematopoietic stem cells from the bone marrow into the circulation, 59,126 as well as movement of immune cells within tissues, which is essential for their function.¹²⁷ In response to stressors, periventricular hypothalamic corticotropin hormone (CRH) neurons stimulate the hypothalamic-pituitary-adrenal axis and regulate trafficking of lymphocytes and monocytes between peripheral tissues and bone marrow.¹²⁸ CRH neurons of the periventricular nucleus and the central nucleus of the amygdala ultimately project to the splenic nerve and can influence adaptive immune responses through both adrenergic and cholinergic mechanisms.⁵⁵ Norepinephrine, released locally from sympathetic nerves or systemically largely from the adrenal gland in a physiological manner or in response to a range of stressors, binds chiefly to the beta2-adreneric receptor (B2AR) on immune cells. Norepinephrine-B2AR signaling can exert immune-suppressive effects such as upregulating PD-1,129 regulating myeloid-derived suppressor cells (MDSCs) and macrophage function and recruitment to tumors, 99,130,131 limiting anti-tumor immunity,^{127,132,133} and promoting T lymphocyte metabolic stress and exhaustion.¹³⁴ Likewise, innervation of solid tumors by sensory neurons can induce T cell exhaustion, preventing effective anti-tumor immunity that could be overcome by inhibiting CGRP, a nociceptor-produced neuropeptide.¹³⁵ Some of the aversive effects of stress on tumor growth in a breast cancer model were shown to be attenuated by







Figure 4. Techniques for studying nervous system-cancer interactions

Methodologies to study nervous system-cancer interactions can be broadly categorized into four dimensions that encompass the functional, structural, and molecular characterization as well as the material or model system that is studied.

optogenetic stimulation of the dopaminergic projections from the ventral tegmental area (VTA) to the medial prefrontal cortex.¹³⁶ Interestingly, VTA activation reduced tumor growth in models of melanoma and lung cancer by modulating the sympathetic innervation to the bone marrow, altering the functional profile of MDSCs.¹³⁷ Together, these results provide a valuable first guidance on how anti-tumor immunotherapies may be augmented by neuromodulation strategies.^{133,138}

Neuronal signaling molecules are sometimes used by the immune system directly. The neurotransmitter GABA can be synthesized by B lymphocytes and, in the context of a mouse model of colon cancer, can bind to GABAA receptors on CD8⁺ T lymphocytes to reduce anti-tumor immunity and enable tumor growth.¹³⁹ Serotonin, secreted by platelets, upregulates PD-L1 expression in models of pancreatic and gastric cancer through histone serotonylation and consequent epigenetic regulation of immune checkpoint expression.¹⁴⁰ In addition, tryptophan, the precursor of serotonin, is metabolized to kynurenines by indoleamine-2,3dioxygenase (IDO) and tryptophan-2,3-dioxygenase (TDO), which are both neuroactive and immunomodulatory, and implicated in neurodegenerative disorders and cancer.¹⁴¹ This also raises the question whether neuronal activity, with secretion of neuronal signaling molecules, can directly influence T lymphocytes and other immune cells. The immunological environment in the CNS is unique, with a very specialized lymphatic system,¹⁴² communication with unique immune populations in the skull bone marrow, and with various neural-immune interactions at the brain borders (for review, see Rustenhoven and Kipnis¹⁴³) that may influence the effectiveness of immunotherapies. Accordingly, neuro-immunooncological interactions are probably quite different within the CNS and outside of it.

The nervous system not only regulates immune responses but also encodes them in an immunological engram in the brain that modifies subsequent immune function. Immune responses to challenges outside of the brain, such as in the gut, can be encoded by neurons in the insular cortex, and reactivation of the neurons activated by a particular immune challenge can recapitulate the immune response operant during the initial immune challenge.¹⁴⁴ This sort of immunological "memory" illustrates that the nervous system exerts profound regulatory control on the immune system, in ways that we are only just beginning to understand.

Another key demonstration of integration between neuronal activity and immune regulation of cancer growth derives from experiments performed using Nf1 optic glioma mice. In addition to light-induced, visual experience-dependent neuronal control of optic glioma initiation and progression. Nf1 mutation in neurons additionally increases basal action potential firing.¹⁰⁹ This increased excitability results in the production of midkine, a paracrine factor of the pleiotrophin family, which acts on T cells to secrete CCL4 and results in microglial secretion of CCL5, a key mitogen for glioma cell growth.^{145,146} Consistent with diverse mechanisms underlying neuronal activity-dependent control of tumor biology, this basal hyperexcitability is mediated by Nf1 protein control of the hyperpolarization-activated cyclic nucleotide-gated potassium channel 1 (HCN1), such that targeting of this channel with the anti-epileptic drug lamotrigine was sufficient to normalize midkine expression and suppress Nf1 optic glioma proliferation in vivo.¹⁰⁹ Moreover, experience-dependent optic nerve activity-regulated shedding of NLGN3⁶⁷ appears to operate through mechanisms distinct from basal Nf1-regulated HCN1-channel-mediated activitydependent expression of midkine. This illustrates how the diversity of neuronal activity-dependent mechanisms may underlie distinct effects on cancer biology in various contexts.¹⁴⁵

Further principles of neuron-immune cell-tumor cell crosstalk are also likely to come to light. How might drugs of neuroscience be leveraged to reduce the immune-suppressive microenvironment of solid tumors and improve immuno-oncology strategies





(Figure 3)? Might targeting neurotransmitter or neuromodulator signaling influence the tumor immune microenvironment to promote anti-tumor immunity? Could targeting growth-promoting interactions between the nervous system and cancer slow down tumor growth to enable immune-based therapies to outpace cancer growth, facilitating tumor regression? Elucidating the mechanisms of nervous system-immune system-cancer interactions may open an important new dimension in immuno-oncology strategies.

Effects of cancer and cancer therapies on the nervous system

Cancer therapies have the potential to limit the very mechanisms of neural homeostasis and plasticity that cancers depend on to grow. Unfortunately, the off-target effects of these therapies on normal neural processes (Figure 1F) can result in a syndrome of debilitating cognitive symptoms characterized by impaired attention, memory, speed of information processing, multitasking and executive function,147 as well as neuropathies affecting sensory, motor, and autonomic peripheral nerves (for review, see Staff et al.¹⁴⁸). Cancer therapies can result in tissue damage within the CNS-particularly insult to white matter and reduced volume of the hippocampus.^{149,150} Furthermore, cancer therapies can disrupt neural communication and network connectivity-which ultimately manifests as cognitive impairment.¹⁵¹⁻¹⁵³ The neurobiological underpinnings of cognitive impairment after cancer therapies (reviewed in Gibson and Monje¹⁵⁴) include radiation- and chemotherapy-induced dysfunction of neural stem and precursor cell populations,^{155–159} dysregulation of hippocampal neurogenesis,^{156,157,160,161} disruption of myelin homeostasis and plasticity,^{32,159} and disruption of synaptic connectivity.162-164

This fundamental understanding has led to therapeutic strategies targeting regeneration of neural stem and precursor cell populations that are showing promise in early clinical studies for cancer therapy-related cognitive impairment.^{165–167} Given how cancers hijack the very same neural mechanisms and structures impaired after traditional cancer therapies, one wonders how the neurotoxicities of cancer therapies contribute to therapeutic efficacy. Understanding this may lead to more specific and less toxic cancer therapeutics.

In addition to neurotoxicity of therapies for cancer, cancer itself can change the nervous system; studies have demonstrated that on a systemic level, mammary gland tumors can disrupt sleep and alter metabolism via altering a specific neuronal population of the CNS.¹⁶⁸ These effects can be observed in cancer patients, who exhibit clinical evidence for behavioral effects of cancer on sleep and appetite.^{169,170} Furthermore, these interactions are bidirectional. On a more local level, tumor-celland neuron-generated paracrine signaling can bidirectionally modulate peripheral sensory nerves, resulting in hypersensitivity, nerve sprouting, and PNI, which contribute to cancer pain.^{171–173} At the level of the whole patient, chronic stress can accelerate metastatic progression of breast and other peripheral cancers by elevating sympathetic signaling.99,174 It will be important to fully understand these bidirectional interactions that seem to constitute a vicious cycle of nervous system-cancer interactions.

A framework for future clinical and preclinical development

Cancer neuroscience is a rapidly evolving field with emerging, exciting discoveries and the potential to influence and even fundamentally change oncological therapies.^{175–177} Furthermore, these discoveries can feed back to inform basic neuroscience and developmental biology. A key challenge is to identify an optimal road to translation for neuroscience-instructed cancer therapies, a path which may be quite different from that of tumor-cell-centric (cytotoxic or molecularly targeted) or anti-tumor immunological strategies. We will discuss trial-enabling aspects and develop a framework for implementing concepts from cancer neuroscience into clinical practice.

An integrative framework that spans diverse preclinical and clinical-translational disciplines will be needed to make progress. In addition to neuroscientific and oncological expertise, development and adaptation of novel technologies will be needed (Figure 4). For both clinical trials and animal studies alike, it will be important to study pharmacological and non-pharmacological interventions over the disease course with a comprehensive clinical characterization, using cancer neuroscience-driven methodological frameworks (Figure 5).

Therapeutic opportunities

A fundamental conviction is that targeting the bidirectional neural-cancer crosstalk can slow tumor growth, or even reverse it, and at the same time preserve or reconstitute quality of life and neurological functioning. Considering the more than one hundred approved drugs in neurology, psychiatry, and internal medicine that interfere with neurotransmitter and other neural signaling, it appears plausible that re-purposing of one or several of those for a given cancer (sub)type and stage can constitute a rapid road for clinical translation (Figure 3). Furthermore, drug development targeting neural-cancer signaling and the functionality of the homotypic and heterotypic nervous system-cancer networks has started, albeit not on a large scale so far.¹⁷⁸ Prospective clinical trials have begun for multiple CNS and systemic cancer types^{175–177}; for an overview of clinical trial numbers, see Pan and Winkler.¹⁷⁹ Some early phase trials have been published,^{180,181} and further results are eagerly awaited. However, interference with the normal function of the CNS and PNS might limit dosing.

Targets and drugs

Figure 3 gives an overview of the principles of cancer neuroscience-related therapies that are tested in distinct indications for various tumor types. Conceptually, the field should prioritize strategies that allow a therapeutic window: since targeted mechanisms of neural-cancer interactions are frequently also relevant for the normal nervous system, a drug concentration needs to be selected that primarily affects cancer biology, or the particular strategy should be localized to affect the tumor microenvironment only (such as denervation strategies of non-brain tumors), always with careful monitoring of CNS and PNS side effects in patients. For children with malignant glioma, an inhibitor of ADAM10/17 (INCB7839) is being tested, because this inhibition blocks the secretion of NLGN3⁶⁹ (NCT04295759). Tumornetwork-disconnecting strategies include the following: (1) the inhibition of gap junctions with meclofenamate in recurrent adult glioblastomas in combination with temozolomide chemotherapy



(MecMeth/NOA-24; EudraCT 2021-000708-39); and (2) targeting glutamatergic neuron-to-glioma synapses with the approved anti-seizure drug perampanel, a non-competitive AMPAR inhibitor, which is underway in a trial initiative in Germany (EudraCT 2023-503938-52).

Outside the CNS, early phase clinical trials have shown that beta-blocker modulation of sympathetic neural signaling is safe in breast cancer patients and well tolerated in combination with neoadjuvant chemotherapy.¹⁸⁰ Findings show that beta-blockers reduce biomarkers of breast cancer cell invasion while improving biomarkers of anti-cancer immunity.^{180,181}

Biomarkers and technologies

It will be crucial to understand whether a given neuroscience-instructed cancer therapy is hitting its target, leading to the desired effects on nervous system-cancer crosstalk, or not. If we do not validate this by accompanying biomarker research, we will not be able to link a positive study result to a desired target engagement. Likewise, we will not be able to interpret a negative result correctly: i.e., was the target pharmacologically missed? Or was it hit, but without a meaningful clinical effect?

Therefore, window-of-opportunity study concepts with investigation of molecular and structural tissue biomarkers of nervous system-cancer interactions in resected or (repetitively) biopsied tumor samples appear particularly meaningful for the first steps,^{180,181} in addition to the development of imaging and electrophysiological biomarkers (Figure 5). Serial investigation over the disease course will enable the study of plasticity and evolution of multicellular neural-cancer networks.

Neural-cancer interactions have been chiefly characterized on a cellular and subcellular level using high-resolution light and electron microscopy as well as electrophysiological patchclamp recordings (Figures 4 and 5). Although these approaches yield a precise readout, it will be difficult to implement these methods on a larger scale for clinical trials. Therefore, a multiomics approach from the macroscopic to the nanoscopic scale (Figure 5) will help define surrogate parameters that can be routinely employed for clinical trials. Additionally, such multiomics approaches will extend our knowledge about the cancer neuroscience-related spatiotemporal cellular as well as molecular heterogeneity and plasticity of cancers. This will require the intensive collaboration of method developers, biologists, clinicians, and biostatisticians. In return, this approach yields the opportunity to methodologically advance not only the cancer neuroscience field but also the neuroscience and oncology fields.

Disease stage

Preclinical work should ideally address the question of whether an anti-cancer therapy that targets neural regulatory mechanisms is likely to be more efficient in the primary setting, in recurrence, or during further metastatic and invasive dissemination, which will define the ideal patient population to include in a trial. Mounting data that particularly resistant and recurrent tumors accumulate neuronal molecular signatures¹⁸² could speak for the latter. On the other hand, secondary evolution of heterogeneity, immune disturbances, and general aspects of patient disposition as well as options for co-treatments and target evaluation, including biological response assessment, favor the newly diagnosed setting.

Outcome parameters

Another important question is the selection of the best efficacy measure, or outcome parameter(s). Targeting conserved neurodevelopmental pathways and structures that have a role in tumor:tumor cell/neuron:tumor cell contacts alike requires careful neurological and neurocognitive as well as behavioral assessments, exceeding the standard batteries in clinical studies. In addition to morphological, physiological, and functional MR imaging and metabolic assessments, network analyses via EEG/ MEG should be considered. Of note, for systemic (non-CNS) cancers, advanced imaging should consider peripheral nerve MRI, which offers a sensitive novel tool for potential effects of the cancer or neuroscience-instructed cancer therapy on sensory nerves.¹⁸³ For any treatment, the first hurdle will be demonstration of a biological impact. This may require tumor or surrogate tissue (CSF/blood)-based diagnostics, i.e., demonstration of change in a preclinically defined biomarker of connectivity or network activity.

Trial design

The primary goal of the early trials should be a definition of the right patient population, which includes stage of the disease (see above), co-treatment as well as target identification, and quantification. Biomarkers from the serum, CSF, and/or tumor tissue, and potentially also imaging biomarkers, may help to identify the patient subpopulation that is most likely to profit from a given neuroscience-instructed cancer therapy, similar to targeted therapy developments in other areas of oncology. As a starting point, phase 0 (window-of-opportunity) trials for neuroscience-instructed cancer therapy appear particularly meaningful, because they include the measurement of drug exposure and biological target engagement in resected tumor tissue. Standardization of clinical protocols will help accelerate the development of effective treatments.

Combination therapy

Therapies targeting neural-cancer interactions might work as monotherapies, but, more likely, they may be used as sensitizers to radiotherapy or chemotherapy (as shown for disconnection strategies of tumor cell networks^{81,88,177}) or to work synergistically to benefit the efficacy and timeline of anti-tumor immuno-therapies, as recently discussed.¹⁸⁴ Targeting neural-cancer interactions may be a required component of effective combination therapy strategies. Therefore, it will be important to devise optimal combination partners, including concomitant cytotoxic, epigenetic, or immunological therapies that together may achieve meaningful clinical effects.

Adapting and developing methodologies for preclinical and basic cancer neuroscience

Further advancing cancer neuroscience, including clinical translation, will require joint efforts in technology development and application in preclinical studies (Figures 4 and 5) that define targetable mechanisms and test novel therapeutic strategies. Using functional imaging techniques such as calcium imaging or voltage imaging would help to decipher functional nervous system-cancer connectivity. Electrophysiological (e.g., microelectrode) arrays can be used to assess electrical connectivity. Ideally, the correlation of functional imaging techniques and spatial transcriptomics would allow for identifying the transcripts that are functionally relevant. The mapping of neuronal input can







Figure 5. Cancer neuroscience from bench-to-bedside and bedside-to-bench

A framework for integrative cancer neuroscience at the intersection of preclinical and translational research. The figure provides an example for brain tumor studies, but it can also serve as a blueprint for extracranial tumors.

be achieved today with the help of evolving, elegant technologies, such as retrograde tracing with advanced viral vectors combined with tissue clearing and light-sheet microsocopy for large-volume imaging.

The analysis of tumor cells in the context of spatial patterns is helpful for the analysis of the tumor microenvironment, and this certainly extends to cancer neuroscience-related questions. For example, it will be important to learn how proximity to neurons and neuronal processes influences cancer cell and tumor immunological features, and whether (how) cancer cell heterogeneity is related to specific neuronal features of cancer cells and/or specific neural interactions. Furthermore, multi-omics strategies that combine the methylome, transcriptome, translatome, and proteome will further increase our knowledge of the molecular machinery associated with the neurobiology of cancer and might reveal novel therapeutic targets.

Finally, methods such as large-scale volume EM and superresolution light microscopy would allow analysis of neuron-tumor connections on a nanoscopic scale. Figure 5 shows a concept of how multi-omics strategies might be integrated across scales for future progress in the field of cancer neuroscience, including clinical and reverse translation.

Mapping the neural-tumor connectome by communitywide efforts

Cancer cell types and their neural partners will need to be classified based on their tumor biological function, connectivity,





physiology, molecular signature, and morphology, in analogy to neuronal cell classification. This will require technological innovation to integrate and understand tumor biological functions. Such initiatives could borrow from neuroscience (e.g., Allen Brain Atlas, neuromorpho.org, and EM connectome data) and oncology (TCGA) consortia to adopt analogous frameworks for cancer neuroscience.

The complexity of such a clinical-translational framework requires a highly interdisciplinary infrastructural framework. Apart from various clinical disciplines that will need to work closely together (e.g., neurology, oncology, neurosurgery, neuropsychology, radiology, and pathology), the close connection to the fields of neuroscience and basic cancer research will be an important element of the collaborative efforts in this direction. Therefore, we believe that establishing and interconnecting specialized cancer neuroscience hubs will be crucial to orchestrate such efforts (Figure 5).

Summary and outlook

Research of the last few years has increasingly consolidated the new field of cancer neuroscience. The demonstrations of direct and indirect influences of the nervous system on cancer initiation, growth, dissemination, and treatment resistance significantly contribute to our understanding of cancer biology today. For every cancer entity investigated so far, cancer-promoting or (less frequently) cancer-inhibitory interactions with the CNS or PNS have been well documented. With more and more mechanisms from more and more cancers reported, the question arises whether nervous system-cancer interactions may someday be regarded as another general principle of cancer pathogenesis. In the next few years, we can expect exciting further discoveries in mechanisms known to be relevant for cancer neuroscience today (Figure 1). In addition, a better understanding of the role of CNS and PNS glial cells and the influence of innervation on other components of the tumor microenvironment will complement our body of knowledge and strengthen the therapeutic armamentarium.

The challenges are clear: we need to better map the nervous system-cancer interactome and connectome on multiple scales and levels. This is a key requirement to gain deeper insight into the complex world of interactions between the nervous system and specific cancer entities and stages. The future selection of the most promising neuroscience-instructed cancer therapies for individual patients will depend on this knowledge, particularly on our ability to conduct meaningful clinical trials and potentially also on feasible biomarkers for nervous system-cancer interactions. Another key requirement for the future will be the joint development of collaborative networks and of cross-disciplinary thinking, methodologies, and research strategies. Cancer neuroscience holds the promise to elucidate fundamentally new and therapeutically important insights into the pathobiology of many—if not all—cancers.

ACKNOWLEDGMENTS

This review article derives from and is written by the participants of a Cancer Neuroscience Think Tank meeting held jointly between the Cancer Neuroscience Programs of Heidelberg University, Stanford University,

and Harvard Medical School on July 18–19, 2022. The authors acknowledge Yvonne Yang for help with Figure 3 design. Initial drafts of figures were made with BioRender.com. The authors are grateful for support from the US National Institutes of Health (DP1NS111132, R01NS092597, P50CA165962, R01CA258384, R01CA261939, R35NS097211, R01DE032018, R37CA242006, U19CA264504), Deutsche Forschungsgemeinschaft (SFB 1389, UNITE Glioblastoma, project ID 404521405, and VE1373/2-1), Else Kroener-Fresenius-Stift (ung (2020-EKEA.135), Virginia and D.K. Ludwig Fund for Cancer Research, ChadTough Defeat DIPG Foundation, Alex's Lemonade Stand Foundation, The University of Texas, MD Anderson Cancer SPORE in Melanoma (P50-CA093459), the Jim Mulva Foundation, and the Sontag Foundation.

AUTHOR CONTRIBUTIONS

M.M., F.W., V.V., and H.S.V. wrote the manuscript; M.M., F.W., and V.V. made the figures; all authors (F.W., H.S.V., M.A., T.B., I.E.D., B.D., D.H.G., S.H.J., T.K., D.M., M.P., A.R., E.K.S., T.C.W., W.W., V.V., and M.M.) edited the manuscript.

DECLARATION OF INTERESTS

M.M. holds equity in MapLight Therapeutics. M.M. and H.S.V. report the patent (US Patent #10,377,818) "Method for treating glioma." F.W. and W.W. report the patent (WO2017020982A1) "Agents for use in the treatment of glioma." F.W. is a co-founder of DC Europa Ltd (a company trading under the name Divide & Conquer) that is developing new medicines for the treatment of glioma. Divide & Conquer also provides research funding to F.W.'s lab under a research collaboration agreement.

REFERENCES

- Boilly, B., Faulkner, S., Jobling, P., and Hondermarck, H. (2017). Nerve dependence: from regeneration to cancer. Cancer Cell 31, 342–354. https://doi.org/10.1016/j.ccell.2017.02.005.
- Kumar, A., and Brockes, J.P. (2012). Nerve dependence in tissue, organ, and appendage regeneration. Trends Neurosci. 35, 691–699. https://doi. org/10.1016/j.tins.2012.08.003.
- Monje, M., Borniger, J.C., D'Silva, N.J., Deneen, B., Dirks, P.B., Fattahi, F., Frenette, P.S., Garzia, L., Gutmann, D.H., Hanahan, D., et al. (2020). Roadmap for the emerging field of cancer neuroscience. Cell *181*, 219– 222. https://doi.org/10.1016/j.cell.2020.03.034.
- Silbereis, J.C., Pochareddy, S., Zhu, Y., Li, M., and Sestan, N. (2016). The cellular and molecular landscapes of the developing human central nervous system. Neuron *89*, 248–268. https://doi.org/10.1016/j.neuron. 2015.12.008.
- Fünfschilling, U., Supplie, L.M., Mahad, D., Boretius, S., Saab, A.S., Edgar, J., Brinkmann, B.G., Kassmann, C.M., Tzvetanova, I.D., Möbius, W., et al. (2012). Glycolytic oligodendrocytes maintain myelin and longterm axonal integrity. Nature 485, 517–521. https://doi.org/10.1038/ nature11007.
- Huxley, A.F., and Stämpeli, R. (1949). Evidence for saltatory conduction in peripheral myelinated nerve fibres. J. Physiol. *108*, 315–339. https:// doi.org/10.1113/jphysiol.1949.sp004335.
- Spitzer, N.C. (2006). Electrical activity in early neuronal development. Nature 444, 707–712. https://doi.org/10.1038/nature05300.
- Bittman, K., Owens, D.F., Kriegstein, A.R., and LoTurco, J.J. (1997). Cell coupling and uncoupling in the ventricular zone of developing neocortex. J. Neurosci. 17, 7037–7044.
- Berg, D.A., Belnoue, L., Song, H., and Simon, A. (2013). Neurotransmitter-mediated control of neurogenesis in the adult vertebrate brain. Development 140, 2548–2561. https://doi.org/10.1242/dev.088005.
- Ohtaka-Maruyama, C., Okamoto, M., Endo, K., Oshima, M., Kaneko, N., Yura, K., Okado, H., Miyata, T., and Maeda, N. (2018). Synaptic transmission from subplate neurons controls radial migration of neocortical neurons. Science 360, 313–317. https://doi.org/10.1126/science.aar2866.





- Ming, G., Henley, J., Tessier-Lavigne, M., Song, H., and Poo, M. (2001). Electrical activity modulates growth cone guidance by diffusible factors. Neuron 29, 441–452. https://doi.org/10.1016/s0896-6273(01)00217-3.
- Catalano, S.M., and Shatz, C.J. (1998). Activity-dependent cortical target selection by thalamic axons. Science 281, 559–562. https://doi.org/10. 1126/science.281.5376.559.
- Dantzker, J.L., and Callaway, E.M. (1998). The development of local, layer-specific visual cortical axons in the absence of extrinsic influences and intrinsic activity. J. Neurosci. 18, 4145–4154.
- Marins, M., Xavier, A.L., Viana, N.B., Fortes, F.S., Fróes, M.M., and Menezes, J.R. (2009). Gap junctions are involved in cell migration in the early postnatal subventricular zone. Dev. Neurobiol. 69, 715–730. https://doi. org/10.1002/dneu.20737.
- Peinado, A., Yuste, R., and Katz, L.C. (1993). Extensive dye coupling between rat neocortical neurons during the period of circuit formation. Neuron 10, 103–114. https://doi.org/10.1016/0896-6273(93)90246-n.
- Picken Bahrey, H.L., and Moody, W.J. (2003). Early development of voltage-gated ion currents and firing properties in neurons of the mouse cerebral cortex. J. Neurophysiol. 89, 1761–1773. https://doi.org/10. 1152/jn.00972.2002.
- Blankenship, A.G., and Feller, M.B. (2010). Mechanisms underlying spontaneous patterned activity in developing neural circuits. Nat. Rev. Neurosci. 11, 18–29. https://doi.org/10.1038/nrn2759.
- Corlew, R., Bosma, M.M., and Moody, W.J. (2004). Spontaneous, synchronous electrical activity in neonatal mouse cortical neurones. J. Physiol. 560, 377–390. https://doi.org/10.1113/jphysiol.2004.071621.
- Bajar, B.T., Phi, N.T., Isaacman-Beck, J., Reichl, J., Randhawa, H., and Akin, O. (2022). A discrete neuronal population coordinates brain-wide developmental activity. Nature 602, 639–646. https://doi.org/10.1038/ s41586-022-04406-9.
- Katz, L.C., and Shatz, C.J. (1996). Synaptic activity and the construction of cortical circuits. Science 274, 1133–1138. https://doi.org/10.1126/science.274.5290.1133.
- Kirkby, L.A., Sack, G.S., Firl, A., and Feller, M.B. (2013). A role for correlated spontaneous activity in the assembly of neural circuits. Neuron 80, 1129–1144. https://doi.org/10.1016/j.neuron.2013.10.030.
- Ojeda, J., and Ávila, A. (2019). Early actions of neurotransmitters during cortex development and maturation of reprogrammed neurons. Front. Synaptic Neurosci. *11*, 33. https://doi.org/10.3389/fnsyn.2019.00033.
- Andäng, M., Hjerling-Leffler, J., Moliner, A., Lundgren, T.K., Castelo-Branco, G., Nanou, E., Pozas, E., Bryja, V., Halliez, S., Nishimaru, H., et al. (2008). Histone H2AX-dependent GABA(A) receptor regulation of stem cell proliferation. Nature *451*, 460–464. https://doi.org/10.1038/ nature06488.
- Gibson, E.M., Purger, D., Mount, C.W., Goldstein, A.K., Lin, G.L., Wood, L.S., Inema, I., Miller, S.E., Bieri, G., Zuchero, J.B., et al. (2014). Neuronal activity promotes oligodendrogenesis and adaptive myelination in the mammalian brain. Science 344, 1252304. https://doi.org/10.1126/science.1252304.
- Paez-Gonzalez, P., Asrican, B., Rodriguez, E., and Kuo, C.T. (2014). Identification of distinct ChAT+ neurons and activity-dependent control of postnatal SVZ neurogenesis. Nat. Neurosci. 17, 934–942. https:// doi.org/10.1038/nn.3734.
- Paul, A., Chaker, Z., and Doetsch, F. (2017). Hypothalamic regulation of regionally distinct adult neural stem cells and neurogenesis. Science 356, 1383–1386. https://doi.org/10.1126/science.aal3839.
- Deisseroth, K., Singla, S., Toda, H., Monje, M., Palmer, T.D., and Malenka, R.C. (2004). Excitation-neurogenesis coupling in adult neural stem/progenitor cells. Neuron 42, 535–552. [S0896627304002661(pii).
- Tozuka, Y., Fukuda, S., Namba, T., Seki, T., and Hisatsune, T. (2005). GABAergic excitation promotes neuronal differentiation in adult hippocampal progenitor cells. Neuron 47, 803–815. https://doi.org/10.1016/j. neuron.2005.08.023.

- Mitew, S., Gobius, I., Fenlon, L.R., McDougall, S.J., Hawkes, D., Xing, Y.L., Bujalka, H., Gundlach, A.L., Richards, L.J., Kilpatrick, T.J., et al. (2018). Pharmacogenetic stimulation of neuronal activity increases myelination in an axon-specific manner. Nat. Commun. *9*, 306. https://doi. org/10.1038/s41467-017-02719-2.
- Hughes, E.G., Orthmann-Murphy, J.L., Langseth, A.J., and Bergles, D.E. (2018). Myelin remodeling through experience-dependent oligodendrogenesis in the adult somatosensory cortex. Nat. Neurosci. 21, 696– 706. https://doi.org/10.1038/s41593-018-0121-5.
- McKenzie, I.A., Ohayon, D., Li, H., de Faria, J.P., Emery, B., Tohyama, K., and Richardson, W.D. (2014). Motor skill learning requires active central myelination. Science 346, 318–322. https://doi.org/10.1126/science. 1254960.
- Geraghty, A.C., Gibson, E.M., Ghanem, R.A., Greene, J.J., Ocampo, A., Goldstein, A.K., Ni, L., Yang, T., Marton, R.M., Paşca, S.P., et al. (2019). Loss of adaptivemyelinationcontributes to methotrexatechemotherapyrelatedcognitiveimpairment. Neuron *103*, 250–265.e8. https://doi.org/ 10.1016/j.neuron.2019.04.032.
- Steadman, P.E., Xia, F., Ahmed, M., Mocle, A.J., Penning, A.R.A., Geraghty, A.C., Steenland, H.W., Monje, M., Josselyn, S.A., and Frankland, P.W. (2020). Disruption of oligodendrogenesisimpairsmemoryconsolidation in adultmice. Neuron *105*, 150–164.e6. https://doi.org/10.1016/j. neuron.2019.10.013.
- Pan, S., Mayoral, S.R., Choi, H.S., Chan, J.R., and Kheirbek, M.A. (2020). Preservation of a remote fear memory requires new myelin formation. Nat. Neurosci. 23, 487–499. https://doi.org/10.1038/s41593-019-0582-1.
- Makinodan, M., Rosen, K.M., Ito, S., and Corfas, G. (2012). A critical period for social experience-dependent oligodendrocyte maturation and myelination. Science 337, 1357–1360. https://doi.org/10.1126/science.1220845.
- Liu, J., Dietz, K., DeLoyht, J.M., Pedre, X., Kelkar, D., Kaur, J., Vialou, V., Lobo, M.K., Dietz, D.M., Nestler, E.J., et al. (2012). Impaired adult myelination in the prefrontal cortex of socially isolated mice. Nat.Neurosci. 15, 1621–1623. https://doi.org/10.1038/nn.3263.
- Yang, S.M., Michel, K., Jokhi, V., Nedivi, E., and Arlotta, P. (2020). Neuron class-specific responses govern adaptive myelin remodeling in the neocortex. Science 370. https://doi.org/10.1126/science.abd2109.
- Pajevic, S., Basser, P.J., and Fields, R.D. (2014). Role of myelin plasticity in oscillations and synchrony of neuronal activity. Neuroscience 276, 135–147. https://doi.org/10.1016/j.neuroscience.2013.11.007.
- Noori, R., Park, D., Griffiths, J.D., Bells, S., Frankland, P.W., Mabbott, D., and Lefebvre, J. (2020). Activity-dependent myelination: A glial mechanism of oscillatory self-organization in large-scale brain networks. Proc. Natl. Acad. Sci. USA *117*, 13227–13237. https://doi.org/10.1073/ pnas.1916646117.
- Bergles, D.E., Roberts, J.D., Somogyi, P., and Jahr, C.E. (2000). Glutamatergic synapses on oligodendrocyte precursor cells in the hippocampus. Nature 405, 187–191. https://doi.org/10.1038/35012083.
- Lin, S.C., and Bergles, D.E. (2004). Synaptic signaling between GABAergic interneurons and oligodendrocyte precursor cells in the hippocampus. Nat. Neurosci. 7, 24–32. https://doi.org/10.1038/nn1162.
- Etxeberria, A., Mangin, J.M., Aguirre, A., and Gallo, V. (2010). Adult-born SVZ progenitors receive transient synapses during remyelination in corpus callosum. Nat. Neurosci. *13*, 287–289. https://doi.org/10.1038/ nn.2500.
- Mount, C.W., Yalçın, B., Cunliffe-Koehler, K., Sundaresh, S., and Monje, M. (2019). Monosynaptic tracing maps brain-wide afferent oligodendrocyte precursor cell connectivity. eLife 8, e49291. https://doi.org/10.7554/ eLife.49291.
- Gautier, H.O., Evans, K.A., Volbracht, K., James, R., Sitnikov, S., Lundgaard, I., James, F., Lao-Peregrin, C., Reynolds, R., Franklin, R.J., et al.





(2015). Neuronal activity regulates remyelination via glutamate signalling to oligodendrocyte progenitors. Nat. Commun. 6, 8518.

- Ortiz, F.C., Habermacher, C., Graciarena, M., Houry, P.Y., Nishiyama, A., Nait Oumesmar, B., and Angulo, M.C. (2019). Neuronal activity in vivo enhances functional myelin repair. JCI Insight 5. https://doi.org/10.1172/jci. insight.123434.
- Zonouzi, M., Scafidi, J., Li, P., McEllin, B., Edwards, J., Dupree, J.L., Harvey, L., Sun, D., Hübner, C.A., Cull-Candy, S.G., et al. (2015). GABAergic regulation of cerebellar NG2 cell development is altered in perinatal white matter injury. Nat. Neurosci. *18*, 674–682. https://doi.org/10.1038/nn.3990.
- Poplawski, G.H.D., Kawaguchi, R., Van Niekerk, E., Lu, P., Mehta, N., Canete, P., Lie, R., Dragatsis, I., Meves, J.M., Zheng, B., et al. (2020). Injured adult neurons regress to an embryonic transcriptional growth state. Nature 581, 77–82. https://doi.org/10.1038/s41586-020-2200-5.
- Kaplan, L., Chow, B.W., and Gu, C. (2020). Neuronal regulation of the blood-brain barrier and neurovascular coupling. Nat. Rev. Neurosci. 21, 416–432. https://doi.org/10.1038/s41583-020-0322-2.
- Krimer, L.S., Muly, E.C., 3rd, Williams, G.V., and Goldman-Rakic, P.S. (1998). Dopaminergic regulation of cerebral cortical microcirculation. Nat. Neurosci. 1, 286–289.
- Pulido, R.S., Munji, R.N., Chan, T.C., Quirk, C.R., Weiner, G.A., Weger, B.D., Rossi, M.J., Elmsaouri, S., Malfavon, M., Deng, A., et al. (2020). Neuronal activityregulatesblood-brainbarriereffluxtransport through endothelialcircadiangenes. Neuron *108*, 937–952.e7. https://doi.org/10. 1016/j.neuron.2020.09.002.
- Kumar, A., Godwin, J.W., Gates, P.B., Garza-Garcia, A.A., and Brockes, J.P. (2007). Molecular basis for the nerve dependence of limb regeneration in an adult vertebrate. Science 318, 772–777. https://doi.org/10. 1126/science.1147710.
- Knox, S.M., Lombaert, I.M., Reed, X., Vitale-Cross, L., Gutkind, J.S., and Hoffman, M.P. (2010). Parasympathetic innervation maintains epithelial progenitor cells during salivary organogenesis. Science 329, 1645– 1647. https://doi.org/10.1126/science.1192046.
- Mahmoud, A.I., O'Meara, C.C., Gemberling, M., Zhao, L., Bryant, D.M., Zheng, R., Gannon, J.B., Cai, L., Choi, W.Y., Egnaczyk, G.F., et al. (2015). Nerves regulate cardiomyocyte proliferation and heart regeneration. Dev. Cell 34, 387–399. https://doi.org/10.1016/j.devcel.2015. 06.017.
- Kreipke, R.E., and Birren, S.J. (2015). Innervating sympathetic neurons regulate heart size and the timing of cardiomyocyte cell cycle withdrawal. J. Physiol. 593, 5057–5073. https://doi.org/10.1113/JP270917.
- Zhang, B., Ma, S., Rachmin, I., He, M., Baral, P., Choi, S., Gonçalves, W.A., Shwartz, Y., Fast, E.M., Su, Y., et al. (2020). Hyperactivation of sympathetic nerves drives depletion of melanocyte stem cells. Nature 577, 676–681. https://doi.org/10.1038/s41586-020-1935-3.
- Brownell, I., Guevara, E., Bai, C.B., Loomis, C.A., and Joyner, A.L. (2011). Nerve-derived sonic hedgehog defines a niche for hair follicle stem cells capable of becoming epidermal stem cells. Cell Stem Cell 8, 552–565. https://doi.org/10.1016/j.stem.2011.02.021.
- Peterson, S.C., Eberl, M., Vagnozzi, A.N., Belkadi, A., Veniaminova, N.A., Verhaegen, M.E., Bichakjian, C.K., Ward, N.L., Dlugosz, A.A., and Wong, S.Y. (2015). Basal cell carcinoma preferentially arises from stem cells within hair follicle and mechanosensory niches. Cell Stem Cell *16*, 400– 412. https://doi.org/10.1016/j.stem.2015.02.006.
- Puzan, M., Hosic, S., Ghio, C., and Koppes, A. (2018). Enteric nervous system regulation of intestinal stem cell differentiation and epithelial monolayer function. Sci. Rep. 8, 6313. https://doi.org/10.1038/s41598-018-24768-3.
- Katayama, Y., Battista, M., Kao, W.M., Hidalgo, A., Peired, A.J., Thomas, S.A., and Frenette, P.S. (2006). Signals from the sympathetic nervous system regulate hematopoieticstem cell egress from bone marrow. Cell 124, 407–421. https://doi.org/10.1016/j.cell.2005.10.041.

- Yamazaki, S., Ema, H., Karlsson, G., Yamaguchi, T., Miyoshi, H., Shioda, S., Taketo, M.M., Karlsson, S., Iwama, A., and Nakauchi, H. (2011). Nonmyelinating Schwann cells maintain hematopoieticstem cell hibernation in the bone marrow niche. Cell *147*, 1146–1158. https://doi.org/10. 1016/j.cell.2011.09.053.
- Knox, S.M., Lombaert, I.M., Haddox, C.L., Abrams, S.R., Cotrim, A., Wilson, A.J., and Hoffman, M.P. (2013). Parasympathetic stimulation improves epithelial organ regeneration. Nat.Commun. *4*, 1494. https://doi.org/10.1038/ncomms2493.
- Schiller, M., Ben-Shaanan, T.L., and Rolls, A. (2021). Neuronal regulation of immunity: why, how and where? Nat. Rev. Immunol. 21, 20–36. https:// doi.org/10.1038/s41577-020-0387-1.
- Carmeliet, P. (2003). Angiogenesis in health and disease. Nat. Med. 9, 653–660. https://doi.org/10.1038/nm0603-653.
- Schiller, M., Azulay-Debby, H., Boshnak, N., Elyahu, Y., Korin, B., Ben-Shaanan, T.L., Koren, T., Krot, M., Hakim, F., and Rolls, A. (2021). Optogenetic activation of local colonic sympathetic innervations attenuates colitis by limiting immune cell extravasation. Immunity 54, 1022–1036.e8. https://doi.org/10.1016/j.immuni.2021.04.007.
- Scherer, H.J. (1938). Structural development in gliomas. Am. J. Cancer 34, 333–351.
- Venkatesh, H.S., Johung, T.B., Caretti, V., Noll, A., Tang, Y., Nagaraja, S., Gibson, E.M., Mount, C.W., Polepalli, J., Mitra, S.S., et al. (2015). Neuronal activitypromotesgliomagrowth through Neuroligin-3 secretion. Cell *161*, 803–816. https://doi.org/10.1016/j.cell.2015.04.012.
- Pan, Y., Hysinger, J.D., Barron, T., Schindler, N.F., Cobb, O., Guo, X., Yalçın, B., Anastasaki, C., Mulinyawe, S.B., Ponnuswami, A., et al. (2021). NF1 mutation drives neuronal activity-dependent initiation of optic glioma. Nature 594, 277–282. https://doi.org/10.1038/s41586-021-03580-6.
- Taylor, K.R., Barron, T., Zhang, H., Hui, A., Hartmann, G., Ni, L., Venkatesh, H.S., Du, P., Mancusi, R., Yalçin, B., et al. (2021). Glioma synapses recruit mechanisms of adaptive plasticity. bioRxiv. https://doi.org/10. 1101/2021.11.04.467325.
- Venkatesh, H.S., Tam, L.T., Woo, P.J., Lennon, J., Nagaraja, S., Gillespie, S.M., Ni, J., Duveau, D.Y., Morris, P.J., Zhao, J.J., et al. (2017). Targeting neuronal activity-regulated neuroligin-3 dependency in high-grade glioma. Nature 549, 533–537. https://doi.org/10.1038/nature24014.
- Chen, P., Wang, W., Liu, R., Lyu, J., Zhang, L., Li, B., Qiu, B., Tian, A., Jiang, W., Ying, H., et al. (2022). Olfactory sensory experience regulates gliomagenesis via neuronal IGF1. Nature 606, 550–556. https://doi.org/ 10.1038/s41586-022-04719-9.
- Venkatesh, H.S., Morishita, W., Geraghty, A.C., Silverbush, D., Gillespie, S.M., Arzt, M., Tam, L.T., Espenel, C., Ponnuswami, A., Ni, L., et al. (2019). Electrical and synaptic integration of glioma into neural circuits. Nature 573, 539–545. https://doi.org/10.1038/s41586-019-1563-y.
- Venkataramani, V., Tanev, D.I., Strahle, C., Studier-Fischer, A., Fankhauser, L., Kessler, T., Körber, C., Kardorff, M., Ratliff, M., Xie, R., et al. (2019). Glutamatergic synaptic input to glioma cells drives brain tumour progression. Nature 573, 532–538. https://doi.org/10.1038/s41586-019-1564-x.
- Venkataramani, V.T.K., Yang, Y., Schubert, M.C., Reyhan, E., Tetzlaff, S.K., Wißmann, N., Botz, M., Soyka, S.J., Beretta, C.A., Pramatarov, R.L., et al. (2022). Glioblastoma hijacks neuronal mechanisms for brain invasion. Cell *185*, 2899–2917.e31. https://doi.org/10.1016/j.cell.2022. 06.054.
- 74. Zeng, Q., Michael, I.P., Zhang, P., Saghafinia, S., Knott, G., Jiao, W., McCabe, B.D., Galván, J.A., Robinson, H.P.C., Zlobec, I., et al. (2019). Synaptic proximity enables NMDAR signalling to promote brain metastasis. Nature 573, 526–531. https://doi.org/10.1038/s41586-019-1576-6.
- Buckingham, S.C., Campbell, S.L., Haas, B.R., Montana, V., Robel, S., Ogunrinu, T., and Sontheimer, H. (2011). Glutamate release by primary



brain tumors induces epileptic activity. Nat.Med. 17, 1269–1274. https://doi.org/10.1038/nm.2453.

- Campbell, S.L., Robel, S., Cuddapah, V.A., Robert, S., Buckingham, S.C., Kahle, K.T., and Sontheimer, H. (2015). GABAergic disinhibition and impaired KCC2 cotransporter activity underlie tumor-associated epilepsy. Glia 63, 23–36. https://doi.org/10.1002/glia.22730.
- John Lin, C.C., Yu, K., Hatcher, A., Huang, T.W., Lee, H.K., Carlson, J., Weston, M.C., Chen, F., Zhang, Y., Zhu, W., et al. (2017). Identification of diverse astrocyte populations and their malignant analogs. Nat. Neurosci. 20, 396–405. https://doi.org/10.1038/nn.4493.
- Yu, K., Lin, C.J., Hatcher, A., Lozzi, B., Kong, K., Huang-Hobbs, E., Cheng, Y.T., Beechar, V.B., Zhu, W., Zhang, Y., et al. (2020). PIK3CA variants selectively initiate brain hyperactivity during gliomagenesis. Nature 578, 166–171. https://doi.org/10.1038/s41586-020-1952-2.
- 79. Krishna, S., Choudhury, A., Seo, K., Ni, L., Kakaizada, S., Lee, A., Aabedi, A., Cao, C., Sudharshan, R., Egladyous, A., et al. (2021). Glioblastoma remodeling of neural circuits in the human brain decreases survival. bioRxiv.
- Numan, T., Breedt, L.C., Maciel, B.A.P.C., Kulik, S.D., Derks, J., Schoonheim, M.M., Klein, M., de Witt Hamer, P.C., Miller, J.J., Gerstner, E.R., et al. (2022). Regional healthy brain activity, glioma occurrence and symptomatology. Brain *145*, 3654–3665. https://doi.org/10.1093/brain/ awac180.
- Osswald, M., Jung, E., Sahm, F., Solecki, G., Venkataramani, V., Blaes, J., Weil, S., Horstmann, H., Wiestler, B., Syed, M., et al. (2015). Brain tumour cells interconnect to a functional and resistant network. Nature 528, 93–98. https://doi.org/10.1038/nature16071.
- Jung, E., Osswald, M., Blaes, J., Wiestler, B., Sahm, F., Schmenger, T., Solecki, G., Deumelandt, K., Kurz, F.T., Xie, R., et al. (2017). Tweety-homolog 1 drivesbraincolonization of gliomas. J. Neurosci. 37, 6837–6850. https://doi.org/10.1523/JNEUROSCI.3532-16.2017.
- Gritsenko, P.G., Atlasy, N., Dieteren, C.E.J., Navis, A.C., Venhuizen, J.H., Veelken, C., Schubert, D., Acker-Palmer, A., Westerman, B.A., Wurdinger, T., et al. (2020). p120-catenin-dependent collective brain infiltration by glioma cell networks. Nat. Cell Biol. 22, 97–107. https://doi.org/ 10.1038/s41556-019-0443-x.
- Nagaraja, S., Quezada, M.A., Gillespie, S.M., Arzt, M., Lennon, J.J., Woo, P.J., Hovestadt, V., Kambhampati, M., Filbin, M.G., Suva, M.L., et al. (2019). Histone variant and cellcontextdetermine H3K27M reprogramming of the enhancerlandscape and oncogenicstate. Mol. Cell 76, 965– 980.e12. https://doi.org/10.1016/j.molcel.2019.08.030.
- Jung, E., Osswald, M., Ratliff, M., Dogan, H., Xie, R., Weil, S., Hoffmann, D.C., Kurz, F.T., Kessler, T., Heiland, S., et al. (2021). Tumor cell plasticity, heterogeneity, and resistance in crucial microenvironmental niches in glioma. Nat.Commun. *12*, 1014. https://doi.org/10.1038/s41467-021-21117-3.
- Linkous, A., Balamatsias, D., Snuderl, M., Edwards, L., Miyaguchi, K., Milner, T., Reich, B., Cohen-Gould, L., Storaska, A., Nakayama, Y., et al. (2019). Modeling patient-derivedglioblastoma with cerebralorganoids. Cell Rep. 26, 3203–3211.e5. https://doi.org/10.1016/j.celrep. 2019.02.063.
- Weil, S., Osswald, M., Solecki, G., Grosch, J., Jung, E., Lemke, D., Ratliff, M., Hänggi, D., Wick, W., and Winkler, F. (2017). Tumor microtubes convey resistance to surgical lesions and chemotherapy in gliomas. Neuro. Oncol. *19*, 1316–1326. https://doi.org/10.1093/neuonc/nox070.
- Schneider, M., Potthoff, A.L., Evert, B.O., Dicks, M., Ehrentraut, D., Dolf, A., Schmidt, E.N.C., Schäfer, N., Borger, V., Pietsch, T., et al. (2021). Inhibition of intercellular cytosolic traffic via gap junctions reinforces lomustine-induced toxicity in glioblastomain dependent of MGMT promoter methylation status. Pharmaceuticals *14*, 195. https://doi.org/10. 3390/ph14030195.
- Le, H.T., Sin, W.C., Lozinsky, S., Bechberger, J., Vega, J.L., Guo, X.Q., Sáez, J.C., and Naus, C.C. (2014). Gap junction intercellular communication mediated by connexin43 in astrocytes is essential for their resistance

to oxidative stress. J. Biol. Chem. 289, 1345–1354. https://doi.org/10. 1074/jbc.M113.508390.

- Chen, Q., Boire, A., Jin, X., Valiente, M., Er, E.E., Lopez-Soto, A., Jacob, L., Patwa, R., Shah, H., Xu, K., et al. (2016). Carcinoma-astrocyte gap junctions promote brain metastasis by cGAMP transfer. Nature 533, 493–498. https://doi.org/10.1038/nature18268.
- Hausmann, D., Hoffmann, D.C., Venkataramani, V., Jung, E., Horschitz, S., Tetzlaff, S.K., Jabali, A., Hai, L., Kessler, T., Azorin, D.D., et al. (2023). Autonomous rhythmic activity in glioma networks drives brain tumour growth. Nature 613, 179–186. https://doi.org/10.1038/s41586-022-05520-4.
- Young, H.H. (1897). On the presence of nerves in tumors and of otherstructures in them as revealed by a modification of Ehrlich's method of "vitalstaining" with methylene blue. J. Exp. Med. 2, 1–12. https://doi. org/10.1084/jem.2.1.1.
- Batsakis, J.G. (1985). Nerves and neurotropic carcinomas. Ann. Otol. Rhinol. Laryngol. 94, 426–427.
- Amit, M., Takahashi, H., Dragomir, M.P., Lindemann, A., Gleber-Netto, F.O., Pickering, C.R., Anfossi, S., Osman, A.A., Cai, Y., Wang, R., et al. (2020). Loss of p53 drives neuron reprogramming in head and neck cancer. Nature 578, 449–454. https://doi.org/10.1038/s41586-020-1996-3.
- Takahashi, T., Ishikura, H., Motohara, T., Okushiba, S., Dohke, M., and Katoh, H. (1997). Perineural invasion by ductal adenocarcinoma of the pancreas. J. Surg. Oncol. 65, 164–170. https://doi.org/10.1002/(sici) 1096-9098(199707)65:3<164::aid-jso4>3.0.co;2-4.
- Villers, A., McNeal, J.E., Redwine, E.A., Freiha, F.S., and Stamey, T.A. (1989). The role of perineural space invasion in the local spread of prostatic adenocarcinoma. J. Urol. *142*, 763–768. https://doi.org/10.1016/ s0022-5347(17)38881-x.
- Zhang, Z., Lei, A., Xu, L., Chen, L., Chen, Y., Zhang, X., Gao, Y., Yang, X., Zhang, M., and Cao, Y. (2017). Similarity in gene-regulatory networks suggests that cancer cells share characteristics of embryonic neural cells. J. Biol. Chem. 292, 12842–12859. https://doi.org/10.1074/jbc. M117.785865.
- Magnon, C., Hall, S.J., Lin, J., Xue, X., Gerber, L., Freedland, S.J., and Frenette, P.S. (2013). Autonomic nerve development contributes to prostate cancer progression. Science 341, 1236361. https://doi.org/10.1126/ science.1236361.
- Sloan, E.K., Priceman, S.J., Cox, B.F., Yu, S., Pimentel, M.A., Tangkanangnukul, V., Arevalo, J.M., Morizono, K., Karanikolas, B.D., Wu, L., et al. (2010). The sympathetic nervous system induces a metastatic switch in primary breast cancer. Cancer Res. 70, 7042–7052. https:// doi.org/10.1158/0008-5472.CAN-10-0522.
- 100. Thaker, P.H., Han, L.Y., Kamat, A.A., Arevalo, J.M., Takahashi, R., Lu, C., Jennings, N.B., Armaiz-Pena, G., Bankson, J.A., Ravoori, M., et al. (2006). Chronic stress promotes tumor growth and angiogenesis in a mouse model of ovarian carcinoma. Nat. Med. *12*, 939–944. https:// doi.org/10.1038/nm1447.
- 101. Hayakawa, Y., Sakitani, K., Konishi, M., Asfaha, S., Niikura, R., Tomita, H., Renz, B.W., Tailor, Y., Macchini, M., Middelhoff, M., et al. (2017). Nerve growth factorpromotesgastrictumorigenesis through aberrantcholinergicsignaling. Cancer Cell *31*, 21–34. https://doi.org/10.1016/j. ccell.2016.11.005.
- 102. Zhao, C.M., Hayakawa, Y., Kodama, Y., Muthupalani, S., Westphalen, C.B., Andersen, G.T., Flatberg, A., Johannessen, H., Friedman, R.A., Renz, B.W., et al. (2014). Denervation suppresses gastric tumorigenesis. Sci. Transl. Med. 6, 250ra115. https://doi.org/10.1126/scitranslmed. 3009569.
- 103. Renz, B.W., Tanaka, T., Sunagawa, M., Takahashi, R., Jiang, Z., Macchini, M., Dantes, Z., Valenti, G., White, R.A., Middelhoff, M.A., et al. (2018). Cholinergic signaling via muscarinic receptors directly and indirectly suppresses pancreatic tumorigenesis and cancer stemness. Cancer Discov. *8*, 1458–1473. https://doi.org/10.1158/2159-8290.CD-18-0046.

Cell Review



- 104. Kamiya, A., Hayama, Y., Kato, S., Shimomura, A., Shimomura, T., Irie, K., Kaneko, R., Yanagawa, Y., Kobayashi, K., and Ochiya, T. (2019). Genetic manipulation of autonomic nerve fiber innervation and activity and its effect on breast cancer progression. Nat.Neurosci. 22, 1289–1305. https:// doi.org/10.1038/s41593-019-0430-3.
- 105. Renz, B.W., Takahashi, R., Tanaka, T., Macchini, M., Hayakawa, Y., Dantes, Z., Maurer, H.C., Chen, X., Jiang, Z., Westphalen, C.B., et al. (2018). beta2 adrenergic-neurotrophin feedforward loop promotes pancreatic cancer. Cancer Cell 33, 75–90.e7. https://doi.org/10.1016/j. ccell.2017.11.007.
- 106. Saloman, J.L., Albers, K.M., Li, D., Hartman, D.J., Crawford, H.C., Muha, E.A., Rhim, A.D., and Davis, B.M. (2016). Ablation of sensory neurons in a genetic model of pancreatic ductal adenocarcinoma slows initiation and progression of cancer. Proc. Natl. Acad. Sci. USA. *113*, 3078–3083. https://doi.org/10.1073/pnas.1512603113.
- 107. Walker, A.K., Martelli, D., Ziegler, A.I., Lambert, G.W., Phillips, S.E., Hill, S.J., McAllen, R.M., and Sloan, E.K. (2019). Circulating epinephrine is not required for chronic stress to enhance metastasis. Psychoneuroendocrinology 99, 191–195. https://doi.org/10.1016/j.psyneuen.2018.09.012.
- Le, C.P., Nowell, C.J., Kim-Fuchs, C., Botteri, E., Hiller, J.G., Ismail, H., Pimentel, M.A., Chai, M.G., Karnezis, T., Rotmensz, N., et al. (2016). Chronic stress in mice remodels lymph vasculature to promote tumour cell dissemination. Nat. Commun. 7, 10634. https://doi.org/10.1038/ ncomms10634.
- 109. Anastasaki, C., Mo, J., Chen, J.K., Chatterjee, J., Pan, Y., Scheaffer, S.M., Cobb, O., Monje, M., Le, L.Q., and Gutmann, D.H. (2022). Neuronal hyperexcitability drives central and peripheral nervous system tumor progression in models of neurofibromatosis-1. Nat. Commun. *13*, 2785. https://doi.org/10.1038/s41467-022-30466-6.
- 110. He, S., Chen, C.H., Chernichenko, N., He, S., Bakst, R.L., Barajas, F., Deborde, S., Allen, P.J., Vakiani, E., Yu, Z., et al. (2014). GFRalpha1 released by nerves enhances cancer cell perineural invasion through GDNF-RET signaling. Proc. Natl. Acad. Sci. USA. *111*, E2008–E2017. https://doi. org/10.1073/pnas.1402944111.
- 111. Ceyhan, G.O., Giese, N.A., Erkan, M., Kerscher, A.G., Wente, M.N., Giese, T., Büchler, M.W., and Friess, H. (2006). The neurotrophic factor artemin promotes pancreatic cancer invasion. Ann. Surg. 244, 274– 281. https://doi.org/10.1097/01.sla.0000217642.68697.55.
- 112. Anand, U., Otto, W.R., Casula, M.A., Day, N.C., Davis, J.B., Bountra, C., Birch, R., and Anand, P. (2006). The effect of neurotrophic factors on morphology, TRPV1 expression and capsaicin responses of cultured human DRG sensory neurons. Neurosci.Lett. 399, 51–56. https://doi.org/ 10.1016/j.neulet.2006.01.046.
- 113. Zhu, Z., Friess, H., diMola, F.F., Zimmermann, A., Graber, H.U., Korc, M., and Büchler, M.W. (1999). Nerve growth factor expression correlates with perineural invasion and pain in human pancreatic cancer. J. Clin. Oncol. 17, 2419–2428. https://doi.org/10.1200/JCO.1999.17.8.2419.
- 114. Au, C.W., Siu, M.K., Liao, X., Wong, E.S., Ngan, H.Y., Tam, K.F., Chan, D.C., Chan, Q.K., and Cheung, A.N. (2009). Tyrosine kinase B receptor and BDNF expression in ovarian cancers - Effect on cell migration, angiogenesis and clinical outcome. Cancer Lett. 281, 151–161. https://doi.org/ 10.1016/j.canlet.2009.02.025.
- Pearse, R.N., Swendeman, S.L., Li, Y., Rafii, D., and Hempstead, B.L. (2005). A neurotrophin axis in myeloma: TrkB and BDNF promote tumor-cell survival. Blood 105, 4429–4436. https://doi.org/10.1182/ blood-2004-08-3096.
- 116. Zahalka, A.H., Arnal-Estapé, A., Maryanovich, M., Nakahara, F., Cruz, C.D., Finley, L.W.S., and Frenette, P.S. (2017). Adrenergic nerves activate an angio-metabolic switch in prostate cancer. Science 358, 321– 326. https://doi.org/10.1126/science.aah5072.
- Banh, R.S., Biancur, D.E., Yamamoto, K., Sohn, A.S.W., Walters, B., Kuljanin, M., Gikandi, A., Wang, H., Mancias, J.D., Schneider, R.J., et al. (2020). Neurons releaseserine to support mRNA translation in pancrea-

ticcancer. Cell 183, 1202–1218.e25. https://doi.org/10.1016/j.cell.2020. 10.016.

- 118. Ayala, G.E., Dai, H., Powell, M., Li, R., Ding, Y., Wheeler, T.M., Shine, D., Kadmon, D., Thompson, T., Miles, B.J., et al. (2008). Cancer-related axonogenesis and neurogenesis in prostate cancer. Clin. Cancer Res. 14, 7593–7603. https://doi.org/10.1158/1078-0432.CCR-08-1164.
- 119. Mauffrey, P., Tchitchek, N., Barroca, V., Bemelmans, A.P., Firlej, V., Allory, Y., Roméo, P.H., and Magnon, C. (2019). Progenitors from the central nervous system drive neurogenesis in cancer. Nature 569, 672– 678. https://doi.org/10.1038/s41586-019-1219-y.
- Payne, S.L., Ram, P., Srinivasan, D.H., Le, T.T., Levin, M., and Oudin, M.J. (2022). Potassium channel-driven bioelectric signalling regulates metastasis in triple-negative breast cancer. EBiomedicine 75, 103767. https://doi.org/10.1016/j.ebiom.2021.103767.
- 121. Rahrmann, E.P., Shorthouse, D., Jassim, A., Hu, L.P., Ortiz, M., Mahler-Araujo, B., Vogel, P., Paez-Ribes, M., Fatemi, A., Hannon, G.J., et al. (2022). The NALCN channel regulates metastasis and nonmalignant cell dissemination. Nat. Genet. 54, 1827–1838. https://doi.org/10.1038/ s41588-022-01182-0.
- 122. Creed, S.J., Le, C.P., Hassan, M., Pon, C.K., Albold, S., Chan, K.T., Berginski, M.E., Huang, Z., Bear, J.E., Lane, J.R., et al. (2015). beta2-adrenoceptor signaling regulates invadopodia formation to enhance tumor cell invasion. Breast Cancer Res. *17*, 145. https://doi.org/10.1186/ s13058-015-0655-3.
- 123. Pascual, G., Domínguez, D., Elosúa-Bayes, M., Beckedorff, F., Laudanna, C., Bigas, C., Douillet, D., Greco, C., Symeonidi, A., Hernández, I., et al. (2021). Dietary palmitic acid promotes a prometastatic memory via Schwann cells. Nature 599, 485–490. https://doi.org/10.1038/ s41586-021-04075-0.
- 124. Borovikova, L.V., Ivanova, S., Zhang, M., Yang, H., Botchkina, G.I., Watkins, L.R., Wang, H., Abumrad, N., Eaton, J.W., and Tracey, K.J. (2000). Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. Nature 405, 458–462. https://doi.org/10.1038/35013070.
- 125. Suzuki, K., Hayano, Y., Nakai, A., Furuta, F., and Noda, M. (2016). Adrenergic control of the adaptive immune response by diurnal lymphocyte recirculation through lymph nodes. J. Exp. Med. 213, 2567–2574. https:// doi.org/10.1084/jem.20160723.
- 126. Méndez-Ferrer, S., Lucas, D., Battista, M., and Frenette, P.S. (2008). Haematopoietic stem cell release is regulated by circadian oscillations. Nature 452, 442–447. https://doi.org/10.1038/nature06685.
- 127. Devi, S., Alexandre, Y.O., Loi, J.K., Gillis, R., Ghazanfari, N., Creed, S.J., Holz, L.E., Shackleford, D., Mackay, L.K., Heath, W.R., et al. (2021). Adrenergic regulation of the vasculature impairs leukocyte interstitial migration and suppresses immune responses. Immunity 54, 1219– 1230.e7. https://doi.org/10.1016/j.immuni.2021.03.025.
- Poller, W.C., Downey, J., Mooslechner, A.A., Khan, N., Li, L., Chan, C.T., McAlpine, C.S., Xu, C., Kahles, F., He, S., et al. (2022). Brain motor and fear circuits regulate leukocytes during acute stress. Nature 607, 578– 584. https://doi.org/10.1038/s41586-022-04890-z.
- 129. Yang, H., Xia, L., Chen, J., Zhang, S., Martin, V., Li, Q., Lin, S., Chen, J., Calmette, J., Lu, M., et al. (2019). Stress-glucocorticoid-TSC22D3 axis compromises therapy-induced antitumor immunity. Nat. Med. 25, 1428–1441. https://doi.org/10.1038/s41591-019-0566-4.
- Mohammadpour, H., MacDonald, C.R., Qiao, G., Chen, M., Dong, B., Hylander, B.L., McCarthy, P.L., Abrams, S.I., and Repasky, E.A. (2019). Beta2 adrenergic receptor-mediated signaling regulates the immunosuppressive potential of myeloid-derived suppressor cells. J. Clin. Invest. *129*, 5537–5552. https://doi.org/10.1172/JCI129502.
- 131. Jiang, S.H., Zhu, L.L., Zhang, M., Li, R.K., Yang, Q., Yan, J.Y., Zhang, C., Yang, J.Y., Dong, F.Y., Dai, M., et al. (2019). GABRP regulates chemokine signalling, macrophage recruitment and tumour progression in pancreatic cancer through tuning KCNN4-mediated Ca2+ signalling in a GABA-independent manner. Gut 68, 1994–2006. https://doi.org/10. 1136/gutjnl-2018-317479.



- 132. Chen, M., Qiao, G., Hylander, B.L., Mohammadpour, H., Wang, X.Y., Subjeck, J.R., Singh, A.K., and Repasky, E.A. (2020). Adrenergic stress constrains the development of anti-tumor immunity and abscopal responses following local radiation. Nat. Commun. *11*, 1821. https://doi. org/10.1038/s41467-020-15676-0.
- Nissen, M.D., Sloan, E.K., and Mattarollo, S.R. (2018). beta-Adrenergic Signaling Impairs antitumor CD8+ T-cell Responses to B-cell Lymphoma Immunotherapy. Cancer Immunol.Res. 6, 98–109. https://doi.org/10. 1158/2326-6066.CIR-17-0401.
- 134. Qiao, G., Chen, M., Mohammadpour, H., MacDonald, C.R., Bucsek, M.J., Hylander, B.L., Barbi, J.J., and Repasky, E.A. (2021). Chronic adrenergicstresscontributes to metabolicdysfunction and an exhaustedphenotype in Tcells in the tumormicroenvironment. Cancer Immunol. Res. 9, 651–664. https://doi.org/10.1158/2326-6066.CIR-20-0445.
- Balood, M., Ahmadi, M., Eichwald, T., Ahmadi, A., Majdoubi, A., Roversi, K., Roversi, K., Lucido, C.T., Restaino, A.C., Huang, S., et al. (2022). Nociceptor neurons affect cancer immunosurveillance. Nature 611, 405–412. https://doi.org/10.1038/s41586-022-05374-w.
- 136. Xu, X.R., Xiao, Q., Hong, Y.C., Liu, Y.H., Liu, Y., and Tu, J. (2021). Activation of dopaminergic VTA inputs to the mPFC ameliorates chronic stressinduced breast tumor progression. CNS Neurosci. Ther. 27, 206–219. https://doi.org/10.1111/cns.13465.
- 137. Ben-Shaanan, T.L., Schiller, M., Azulay-Debby, H., Korin, B., Boshnak, N., Koren, T., Krot, M., Shakya, J., Rahat, M.A., Hakim, F., et al. (2018). Modulation of anti-tumor immunity by the brain's reward system. Nat.Commun. 9, 2723. https://doi.org/10.1038/s41467-018-05283-5.
- 138. Kokolus, K.M., Zhang, Y., Sivik, J.M., Schmeck, C., Zhu, J., Repasky, E.A., Drabick, J.J., and Schell, T.D. (2018). Beta blocker use correlates with better overall survival in metastatic melanoma patients and improves the efficacy of immunotherapies in mice. Oncoimmunology 7, e1405205. https://doi.org/10.1080/2162402X.2017.1405205.
- 139. Zhang, B., Vogelzang, A., Miyajima, M., Sugiura, Y., Wu, Y., Chamoto, K., Nakano, R., Hatae, R., Menzies, R.J., Sonomura, K., et al. (2021). B cellderived GABA elicits IL-10+macrophages to limit anti-tumour immunity. Nature 599, 471–476. https://doi.org/10.1038/s41586-021-04082-1.
- 140. Schneider, M.A., Heeb, L., Beffinger, M.M., Pantelyushin, S., Linecker, M., Roth, L., Lehmann, K., Ungethüm, U., Kobold, S., Graf, R., et al. (2021). Attenuation of peripheral serotonin inhibits tumor growth and enhances immune checkpoint blockade therapy in murine tumor models. Sci. Transl. Med. *13*, eabc8188. https://doi.org/10.1126/scitranslmed. abc8188.
- 141. Platten, M., Nollen, E.A.A., Röhrig, U.F., Fallarino, F., and Opitz, C.A. (2019). Tryptophan metabolism as a common therapeutic target in cancer, neurodegeneration and beyond. Nat. Rev. Drug Discov. 18, 379– 401. https://doi.org/10.1038/s41573-019-0016-5.
- 142. Louveau, A., Smirnov, I., Keyes, T.J., Eccles, J.D., Rouhani, S.J., Peske, J.D., Derecki, N.C., Castle, D., Mandell, J.W., Lee, K.S., et al. (2015). Structural and functional features of central nervous system lymphatic vessels. Nature 523, 337–341. https://doi.org/10.1038/nature14432.
- 143. Rustenhoven, J., and Kipnis, J. (2022). Brain borders at the central stage of neuroimmunology. Nature 612, 417–429. https://doi.org/10.1038/ s41586-022-05474-7.
- 144. Koren, T., Yifa, R., Amer, M., Krot, M., Boshnak, N., Ben-Shaanan, T.L., Azulay-Debby, H., Zalayat, I., Avishai, E., Hajjo, H., et al. (2021). Insular cortex neurons encode and retrieve specific immune responses. Cell 184, 5902–5915.e17. https://doi.org/10.1016/j.cell.2021.10.013.
- 145. Guo, X., Pan, Y., Xiong, M., Sanapala, S., Anastasaki, C., Cobb, O., Dahiya, S., and Gutmann, D.H. (2020). Midkine activation of CD8+ Tcells establishes a neuron-immune-cancer axis responsible for low-grade glioma growth. Nat. Commun. *11*, 2177. https://doi.org/10.1038/s41467-020-15770-3.
- 146. SOlga, A.C., Pong, W.W., Kim, K.Y., Cimino, P.J., Toonen, J.A., Walker, J., Wylie, T., Magrini, V., Griffith, M., Griffith, O.L., et al. (2015). RNA sequencing of tumor-associatedmicrogliareveals Ccl5 as a stromalche-

mokinecritical for Neurofibromatosis-1 gliomagrowth. Neoplasia 17, 776–788. https://doi.org/10.1016/j.neo.2015.10.002.

- 147. Wefel, J.S., Kayl, A.E., and Meyers, C.A. (2004). Neuropsychological dysfunction associated with cancer and cancer therapies: a conceptual review of an emerging target. Br. J. Cancer 90, 1691–1696. https://doi. org/10.1038/sj.bjc.6601772.
- 148. Staff, N.P., Grisold, A., Grisold, W., and Windebank, A.J. (2017). Chemotherapy-induced peripheral neuropathy: A current review. Ann. Neurol. *81*, 772–781. https://doi.org/10.1002/ana.24951.
- Kesler, S.R., Sleurs, C., McDonald, B.C., Deprez, S., van der Plas, E., and Nieman, B.J. (2021). Brain imaging in pediatriccancersurvivors: correlates of cognitiveimpairment. J. Clin. Oncol. 39, 1775–1785. https://doi. org/10.1200/JCO.20.02315.
- McDonald, B.C. (2021). Structural neuroimagingfindingsrelated to adultnon-CNS cancer and treatment: Review, integration, and implications for treatment of cognitive dysfunction: Review. Neurotherapeutics 18, 792– 810. https://doi.org/10.1007/s13311-021-01096-5.
- 151. Bells, S., Lefebvre, J., Prescott, S.A., Dockstader, C., Bouffet, E., Skocic, J., Laughlin, S., and Mabbott, D.J. (2017). Changes in white mattermicrostructureimpactcognition by disrupting the ability of neuralassemblies to synchronize. J. Neurosci. 37, 8227–8238. https://doi.org/10.1523/ JNEUROSCI.0560-17.2017.
- 152. Oyefiade, A., Moxon-Emre, I., Beera, K., Bouffet, E., Taylor, M., Ramaswamy, V., Laughlin, S., Skocic, J., and Mabbott, D. (2022). Structural connectivity and intelligence in brain-injured children. Neuropsychologia *173*, 108285. https://doi.org/10.1016/j.neuropsychologia.2022.108285.
- 153. Gauvreau, S., Lefebvre, J., Bells, S., Laughlin, S., Bouffet, E., and Mabbott, D.J. (2019). Disrupted network connectivity in pediatric brain tumor survivors is a signature of injury. J. Comp. Neurol. 527, 2896–2909. https://doi.org/10.1002/cne.24717.
- Gibson, E.M., and Monje, M. (2021). Microglia in cancertherapy-relatedcognitiveimpairment. Trends Neurosci. 44, 441–451. https://doi.org/10. 1016/j.tins.2021.02.003.
- 155. Parent, J.M., Tada, E., Fike, J.R., and Lowenstein, D.H. (1999). Inhibition of dentate granule cell neurogenesis with brain irradiation does not prevent seizure-induced mossy fiber synaptic reorganization in the rat. J. Neurosci. 19, 4508–4519.
- Monje, M.L., Mizumatsu, S., Fike, J.R., and Palmer, T.D. (2002). Irradiation induces neural precursor-cell dysfunction. Nat. Med. 8, 955–962. https://doi.org/10.1038/nm749.
- 157. Monje, M.L., Toda, H., and Palmer, T.D. (2003). Inflammatory blockade restores adult hippocampal neurogenesis. Science 302, 1760–1765. https://doi.org/10.1126/science.10884171088417.
- Dietrich, J., Han, R., Yang, Y., Mayer-Pröschel, M., and Noble, M. (2006). CNS progenitor cells and oligodendrocytes are targets of chemotherapeutic agents in vitro and in vivo. J. Biol. 5, 22. https://doi.org/10.1186/ jbiol50.
- 159. Gibson, E.M., Nagaraja, S., Ocampo, A., Tam, L.T., Wood, L.S., Pallegar, P.N., Greene, J.J., Geraghty, A.C., Goldstein, A.K., Ni, L., et al. (2019). Methotrexate chemotherapy induces persistent tri-glial dysregulation that underlies chemotherapy-related cognitive impairment. Cell *176*, 43–55.e13. https://doi.org/10.1016/j.cell.2018.10.049.
- Monje, M.L., Vogel, H., Masek, M., Ligon, K.L., Fisher, P.G., and Palmer, T.D. (2007). Impaired human hippocampal neurogenesis after treatment for central nervous system malignancies. Ann.Neurol. 62, 515–520. https://doi.org/10.1002/ana.21214.
- 161. Seigers, R., Schagen, S.B., Coppens, C.M., van der Most, P.J., van Dam, F.S., Koolhaas, J.M., and Buwalda, B. (2009). Methotrexate decreases hippocampal cell proliferation and induces memory deficits in rats. Behav. Brain Res. 201, 279–284. https://doi.org/10.1016/j.bbr.2009. 02.025.
- 162. Andres, A.L., Gong, X., Di, K., and Bota, D.A. (2014). Low-doses of cisplatin injure hippocampal synapses: a mechanism for 'chemo' brain?





Exp. Neurol. 255, 137–144. https://doi.org/10.1016/j.expneurol.2014. 02.020.

- Parihar, V.K., and Limoli, C.L. (2013). Cranial irradiation compromises neuronal architecture in the hippocampus. Proc. Natl. Acad. Sci. USA. *110*, 12822–12827. https://doi.org/10.1073/pnas.1307301110.
- 164. Kempf, S.J., Buratovic, S., von Toerne, C., Moertl, S., Stenerlöw, B., Hauck, S.M., Atkinson, M.J., Eriksson, P., and Tapio, S. (2014). Ionising radiation immediately impairs synaptic plasticity-associated cytoskeletal signalling pathways in HT22 cells and in mouse brain: an in vitro/in vivo comparison study. PLoS One 9, e110464. https://doi.org/10.1371/journal.pone.0110464.
- 165. Ayoub, R., Ruddy, R.M., Cox, E., Oyefiade, A., Derkach, D., Laughlin, S., Ades-Aron, B., Shirzadi, Z., Fieremans, E., MacIntosh, B.J., et al. (2020). Assessment of cognitive and neural recovery in survivors of pediatric brain tumors in a pilot clinical trial using metformin. Nat.Med. 26, 1285– 1294. https://doi.org/10.1038/s41591-020-0985-2.
- 166. Mabbott, D.J., Monsalves, E., Spiegler, B.J., Bartels, U., Janzen, L., Guger, S., Laperriere, N., Andrews, N., and Bouffet, E. (2011). Longitudinal evaluation of neurocognitive function after treatment for central nervous system germ cell tumors in childhood. Cancer *117*, 5402–5411. https://doi.org/10.1002/cncr.26127.
- 167. Riggs, L., Piscione, J., Laughlin, S., Cunningham, T., Timmons, B.W., Courneya, K.S., Bartels, U., Skocic, J., de Medeiros, C., Liu, F., et al. (2017). Exercise training for neural recovery in a restricted sample of pediatric brain tumor survivors: a controlled clinical trial with crossover of training versus no training. Neuro. Oncol. 19, 440–450. https://doi.org/ 10.1093/neuonc/now177.
- 168. Borniger, J.C., Walker Ii, W.H., Surbhi Emmer, K.M., Zhang, N., Zalenski, A.A., Muscarella, S.L., Fitzgerald, J.A., Smith, A.N., Braam, C.J., et al. (2018). A role for hypocretin/orexin in metabolic and sleep abnormalities in a mouse model of non-metastatic breast cancer. Cell Metab 28, 118– 129. https://doi.org/10.1016/j.cmet.2018.04.021.
- 169. Palesh, O., Aldridge-Gerry, A., Zeitzer, J.M., Koopman, C., Neri, E., Giese-Davis, J., Jo, B., Kraemer, H., Nouriani, B., and Spiegel, D. (2014). Actigraphy-measured sleep disruption as a predictor of survival among women with advanced breast cancer. Sleep *37*, 837–842. https://doi. org/10.5665/sleep.3642.
- 170. Ezeoke, C.C., and Morley, J.E. (2015). Pathophysiology of anorexia in the cancer cachexia syndrome. J. Cachexia Sarcopenia Muscle 6, 287–302. https://doi.org/10.1002/jcsm.12059.
- 171. Schweizerhof, M., Stösser, S., Kurejova, M., Njoo, C., Gangadharan, V., Agarwal, N., Schmelz, M., Bali, K.K., Michalski, C.W., Brugger, S., et al. (2009). Hematopoieticcolony-stimulating factors mediate tumor-nerve interactions and bone cancer pain. Nat. Med. 15, 802–807. https://doi. org/10.1038/nm.1976.
- 172. Selvaraj, D., Gangadharan, V., Michalski, C.W., Kurejova, M., Stösser, S., Srivastava, K., Schweizerhof, M., Waltenberger, J., Ferrara, N., Heppenstall, P., et al. (2015). A functionalrole for VEGFR1 expressed in peripheralsensoryneurons in cancerpain. Cancer Cell 27, 780–796. https://doi. org/10.1016/j.ccell.2015.04.017.
- 173. Hirth, M., Gandla, J., Höper, C., Gaida, M.M., Agarwal, N., Simonetti, M., Demir, A., Xie, Y., Weiss, C., Michalski, C.W., et al. (2020). CXCL10 and CCL21 promotemigration of pancreatic cancer cells toward sensory

neurons and neural remodeling in tumors in mice, associated with pain in patients. Gastroenterology *159*, 665–681.e13. https://doi.org/10. 1053/j.gastro.2020.04.037.

- 174. Moreno-Smith, M., Lutgendorf, S.K., and Sood, A.K. (2010). Impact of stress on cancer metastasis. Future Oncol. 6, 1863–1881. https://doi. org/10.2217/fon.10.142.
- 175. Shi, D.D., Guo, J.A., Hoffman, H.I., Su, J., Mino-Kenudson, M., Barth, J.L., Schenkel, J.M., Loeffler, J.S., Shih, H.A., Hong, T.S., et al. (2022). Therapeutic avenues for cancer neuroscience: translational frontiers and clinical opportunities. Lancet Oncol. 23, e62–e74. https://doi.org/ 10.1016/S1470-2045(21)00596-9.
- 176. Demir, I.E., Mota Reyes, C., Alrawashdeh, W., Ceyhan, G.O., Deborde, S., Friess, H., Görgülü, K., Istvanffy, R., Jungwirth, D., Kuner, R., et al. (2021). Future directions in preclinical and translational cancer neuroscience research. Nat. Cancer *1*, 1027–1031. https://doi.org/10.1038/ s43018-020-00146-9.
- 177. Venkataramani, V., Schneider, M., Giordano, F.A., Kuner, T., Wick, W., Herrlinger, U., and Winkler, F. (2022). Disconnecting multicellular networks in brain tumours. Nat. Rev. Cancer 22, 481–491. https://doi.org/ 10.1038/s41568-022-00475-0.
- Dolgin, E. (2020). Cancer-neuronal crosstalk and the startups working to silence it. Nat. Biotechnol. 38, 115–117. https://doi.org/10.1038/s41587-020-0411-9.
- 179. Pan, C., and Winkler, F. (2022). Insights and opportunities at the crossroads of cancer and neuroscience. Nat. Cell Biol. 24, 1454–1460. https://doi.org/10.1038/s41556-022-00978-w.
- 180. Hiller, J.G., Cole, S.W., Crone, E.M., Byrne, D.J., Shackleford, D.M., Pang, J.B., Henderson, M.A., Nightingale, S.S., Ho, K.M., Myles, P.S., et al. (2020). Preoperative beta-blockade with propranololreducesbiomarkers of metastasis in breastcancer: A Phase II randomized trial. Clin. Cancer Res. 26, 1803–1811. https://doi.org/10.1158/1078-0432. CCR-19-2641.
- 181. Shaashua, L., Shabat-Simon, M., Haldar, R., Matzner, P., Zmora, O., Shabtai, M., Sharon, E., Allweis, T., Barshack, I., Hayman, L., et al. (2017). Perioperative COX-2 and beta-adrenergic blockade improves metastatic biomarkers in breast cancer patients in a Phase-II randomized trial. Clin. Cancer Res. 23, 4651–4661. https://doi.org/10.1158/1078-0432.CCR-17-0152.
- 182. Varn, F.S., Johnson, K.C., Martinek, J., Huse, J.T., Nasrallah, M.P., Wesseling, P., Cooper, L.A.D., Malta, T.M., Wade, T.E., Sabedot, T.S., et al. (2022). Glioma progression is shaped by genetic evolution and microenvironment interactions. Cell *185*, 2184–2199.e16. https://doi.org/10. 1016/j.cell.2022.04.038.
- 183. Jende, J.M.E., Groener, J.B., Kender, Z., Rother, C., Hahn, A., Hilgenfeld, T., Juerchott, A., Preisner, F., Heiland, S., Kopf, S., et al. (2020). Structural nerveremodeling at 3-T MR neurographydiffers between painful and painlessdiabeticpolyneuropathy in Type 1 or 2 diabetes. Radiology 294, 405–414. https://doi.org/10.1148/radiol.2019191347.
- 184. Majzner, R.G., Ramakrishna, S., Yeom, K.W., Patel, S., Chinnasamy, H., Schultz, L.M., Richards, R.M., Jiang, L., Barsan, V., Mancusi, R., et al. (2022). GD2-CAR T cell therapy for H3K27M-mutated diffuse midline gliomas. Nature 603, 934–941. https://doi.org/10.1038/s41586-022-04489-4.